



**4FCROPS**

**Future Crops for  
Food, Feed, Fiber and Fuel**

# **Work Package 2**

## **Cropping Possibilities**

### **Task 2.5**

## **Biotechnological Improvement**

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# INTRODUCTION

Genomics and biotechnology are the modern tools for understanding plants at the various biological and environmental levels, as well as boosting classical plant breeding techniques. The application of new biotech technologies in plant breeding can help provide a more sustainable solution in view of the fact that we are facing a conflict between rising demand and environmental concerns. Advances in plant breeding methods could help boost non-food crops' yield, quality and adaptation to various biotic and abiotic factors (heat, cold, pathogens, water), while reducing the cost of production.

Plants and plant derived materials hold great potential to provide renewable products for the future. In forthcoming decades, the utilisation of crops for energy productions is expected to increase. Biofuels can be produced from a variety of biomass types which are the main source (65%) of renewable energy in EU25 ([http://ec.europa.eu/research/energy/pdf/biomass\\_en.pdf](http://ec.europa.eu/research/energy/pdf/biomass_en.pdf)).

The objective of Task 2.5 was to review the current status for selected non-food crops by the 4FCrops consortium, summarizing aspects of their genetics, genomics and breeding. Nowadays, the available literature forms the basis for current and future efforts to introduce and establish the selected non-food crops, along with strategies to produce new genetic material for biofuel, feed and fiber exploitation.

The current report intends to assess the capacity of biotechnological applications to develop a beneficial pipeline extending from feedstock development to sustainable biomaterials/biofuel production and provide examples of the current state-of-the-art on future non-food crops.

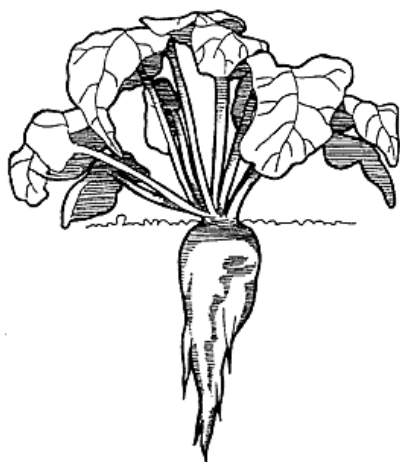
# SUGAR-PRODUCING CROPS

## *Beta Vulgaris L.*

### Introduction

Sugar beet (*Beta vulgaris* L.), a member of the delicious food group of the *Chenopodiaceae* family, is a plant whose root contains a high concentration of sucrose. It is grown commercially for sugar production. The European Union, the United States, and Russia are the world's three largest sugar beet producers, although only the European Union and Ukraine are significant exporters of sugar from beets. The U.S. harvested 1,004,600 acres of sugar beets in 2008 alone. Beet sugar accounts for 30% of the world's sugar production.

Sugar beet is a hardy biennial plant that can be grown commercially in a wide variety of temperate climates. During its first growing season, it produces a large (1–2 kg) storage root whose dry mass is 15–20% sucrose by weight. If the plant is not harvested at this time, then during its second growing season, nutrients in the root will be used to produce flowers and seeds and the root will decrease in size. In commercial beet production, the root is harvested after the first growing season.



**Figure 1:** A mature sugar beet plant

### Scientific classification

Kingdom:	Plantae
Division:	Magnoliophyta
Class:	Magnoliopsida
Order:	Caryophyllales
Family:	Amaranthaceae
Subfamily:	Chenopodiaceae
Genus:	<i>Beta</i>
Species:	<i>B. vulgaris</i>

### Biotechnology Approaches

#### Germination studies

There have been studies in order to investigate sugar beet germination. In the recent years, many investigators have shown that crucial role during the germination period play the

plant hormones ABA (Abscisic acid) and ACC (precursor of ethylene) (Hermann et al., 2007). Molecular approaches used by research groups revealed a set of key regulator genes involved in sugar beet's abiotic stress responses (McGrath et al., 2008). In addition, there have been identified proteins involved specific in germination (de los Reyes, 2003) and in the production of the different parts of the plant such as root, cotyledons and perisperm (Catusse et al., 2008).

### **Flowering**

The sugar beet and its wild progenitor sea beet (*Beta vulgaris ssp. maritima*) are facultative perennials that, under natural growing conditions, exhibit either an annual or a biennial flowering behavior. Many researchers identified that the difference in flowering phenology between annual and biennial sugar beets is determined by a single gene known as the "bolting gene" (Abe et al., 1997).

The effects of the environment on floral initiation in sugar beet have been observed for years but the molecular mechanisms that underlie its control remain to be fully elucidated. As mentioned above, the transition is signified by rapid elongation of the stem (bolting) and tightly linked to the dominant early-bolting (Bolting) gene. In winter-annual ecotypes of *Arabidopsis thaliana*, expression of the MADS-box transcription factor FLOWERING LOCUS C (FLC) during the first growing season creates a facultative vernalization requirement. Reeves et al, (2007) identified FLC homologs in sugar beet by querying expressed sequence tag (EST) databases from *B. vulgaris*.

### **Molecular markers**

Species of the genus *Beta* are grouped into four sections *Beta*, *Corollinae*, *Nanae* and *Procumbentes*. All cultivated beets (sugar, fodder, garden and leaf beet) belong exclusively to the section *Beta*. With approx. 20 closely and distantly related species and subspecies, the genus provides a suitable system for the comparative study of nuclear genome composition and evolution. Many genus-, section- or species specific repetitive DNA sequences have been analysed from cultivated and wild *Beta* species (Kubis et al., 1998; Gao et al., 2000), with the most widely used to be the structural modifications of the centromere-specific satellites (Dechyeva et al., 2003; Menzel et al., 2008).

### **Breeding**

Sugar beet is an important agricultural crop, and the results of genome research in this species might be important to the practical implementation in green biotechnology. Currently, a fine-resolution physical map is under construction and a genome-sequencing project is carried out in the framework GABI–Genome Analysis in Biological System Plant (<http://www.gabi.de/>) aiming to unravel the genome composition of this crop species. Interspecific hybrids and addition lines of *B. vulgaris* are a valuable starting material for plant breeders and an interesting object for fundamental studies on plant genome composition and evolution (Jacobs et al., 2009). Cytoplasmic male sterility (CMS), the maternally inherited

failure to produce functional pollen, has also used in the breeding of sugar beet (Satoh et al. 2004; Yamamoto et al., 2005).

### **Genetic modification**

Transgenic approaches aim to two directions: towards modification of specific traits comprising the increase of pathogen resistance, sugar content and improvement of sugar storage (Graham et al., 1997; Zhang et al., 2000) and towards the tissue specific high protein production in the transgenic plants (Outchkourov et al., 2003; Jaeger et al., 2002).

Disease control is one of the most important goals for biotechnological approaches towards improving sugar beet performance. There are many leaf spot diseases that are detrimental to the plant with the most widespread, destructive but also studied been the *Cercospora* leaf spot (Stahl et al., 2004).

In order to achieve the production of these transgenic lines, *Beta Vulgaris* plants are transformed via the *Agrobacterium tumefaciens* transformation technique which is being utilised since 1991 for this plant species (Lindsey et al., 1991; Stahl et al., 2004) and via the biolistic transformation method (Stahl et al., 2004).

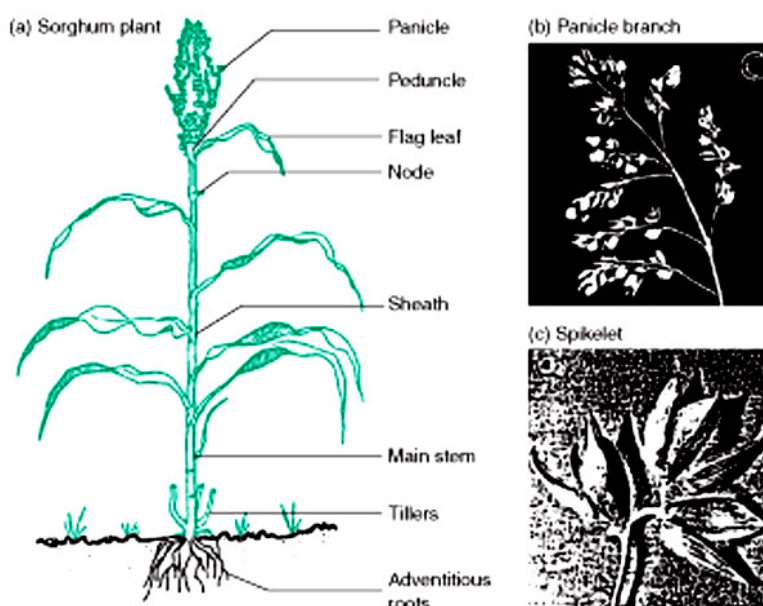
## ***Sorghum bicolor* L.**

### **Introduction**

Sorghum is a single- to multi-culmed C4 plant with perfect flowers; grass species cultivated in diverse and adverse environments from subhumid, hot and dry agro-ecologies, to drought-prone low-to-medium altitudes of the tropics and subtropical regions of the world. This very versatile crop is truly multipurpose, and is used as: (1) grain for food, livestock feed, and industrial products like malt, alcoholic and nonalcoholic beverages, lager beer, stout, and malt drinks; (2) crop residue and silage for livestock feed; (3) chewing cane of the sweet stalk sorghums, (4) household appliances (in fencing and roofing with the dried stalks and as a broom for sweeping with the broomcorn types); and (5) sources of industrial alcohol and household brown sugar with the sweetstalk sorghums. The sorghum plant is composed of two major sections: (1) the vegetative part consisting of the fibrous root system, the culm (stem), and leaves with leaf sheaths wrapping around the node and internode of the culm; and (2) the reproductive portion called inflorescence (panicle) carried on a peduncle (neck, which can be straight or curved (goose neck)) which can be well exerted (short or long neck) or poorly exerted with panicle partially covered by the boot (flag leaf and sheath). The peduncle extends into a central axis of the panicle called rachis, from the nodes of which several branches originate, which bear racemes.

In the preserved germplasm accessions, the unique diversity of sorghum has been classified according to species. There are the cultivated sorghum (*Sorghum bicolor* L. Moench) and the wild weedy species. Within cultivated sorghums, there are five basic races (race guinea, race candatum, race durra, race bicolor, and race kafir) and ten stable hybrid races.

The taxonomy and evolution of sorghum is well known and documented. The cultivated sorghum, *S. bicolor* (L. Moench) is made up of two crosscompatible subspecies, bicolor and arundinaceum. Bicolor is derived from the domestication of the wild and weedy *Sorghum arundinaceum*.



**Figure 1:** Diagram of the sorghum plant (a) and its components (b and c).

### Biotechnology approaches

In recent times, more nonconventional breeding approaches have been used to improve sorghum. These include molecular breeding, biotechnological approaches, and farmer participatory plant breeding (PPB). Each of these has their specific uses, which can overlap in some instances in their complementarity with, and enhancing research in conventional breeding. In sorghum, biotechnology tools are now being used in drought-resistance breeding by tagging quantitative trait loci (QTLs) associated with the different types of drought resistance (seedling, pre-flowering, and postflowering stages), for Striga resistance, and genetic mapping for linkage and genomic maps. They are also used for stem borer and midge resistance, grain quality improvement for increased protein, better digestibility, better processing, and incorporation of vitamin A (enhancing yellow endosperm sorghums) and micronutrients (especially Fe, Zn, and Ca); fodder and crop residue quality and digestibility by

incorporation of bmr gene for brown midrib and stay-green trait gene. Newer biotechnological techniques have recently been used in developing alternative foods and industrial applications (as in using sorghum nondigestible protein character in developing biofilms for fruits and vegetable preservation for exports). These several methodologies have been described and recorded, especially for striga resistance and control, herbicide resistance, molecular and linkage mapping, population dynamics of striga, and biodiversity studies in wide crosses.

### **Genetic screens**

Motivated by interest in a range of basic and applied questions, the linkage maps of sorghum have been employed in the “tagging” (mapping) of genes for a large number of traits. The interspecific population has been especially useful for characterization of genes related to domestication, such as seed size, shattering (Paterson et al, 1995), tillering, and rhizomatousness (Paterson et al, 1995). Plant height and flowering time (Ulanich et al, 1996) have been a high priority. Similarly, the importance of hybrid sorghum motivated much research into the genetic control of fertility restoration (Wen et al, 2002). Resistance genes have been tagged for numerous diseases, key insect pests, and also the parasitic weed, striga (Mutengwa et al, 2005). Genes and QTLs have been identified that are related to abiotic stresses including postreproductive stage drought tolerance (stay-green); preharvest sprouting (Carrari et al, 2003), and aluminum tolerance (Magalhaes et al, 2004). Additional morphological characteristics have also been mapped in interspecific and/or intraspecific populations (Feltus et al, 2006). As with most agronomically important crop species, sorghum genomics is an area of active research. In the past 10 years, over 11 genetic linkage maps of sorghum have been published (McIntyre et al., 2001). Sorghum transformation has been reported and repeated by several research groups, both public and private and it is now possible to create transgenic sorghum.

Compared with other cereal grains, *Sorghum bicolor* shows lower protein digestibility. The low digestibility is thought to result from disulfide cross linking in the  $\beta$ - and  $\gamma$ -kafirins (Duodu et al, 2003). In contrast, the single recessive high digestibility/high lysine content (HD) mutation which confers greater grain digestibility exists in sorghum that is thought to result from reduced accumulation of  $\gamma$ - kafirin that allows greater access to the high digestible  $\alpha$ -kafarin fraction. In an effort to both clearly define the molecular basis for the HD trait and develop tools to improve the introgression of this difficult-to-screen trait, Winn et al (2009) focused on mapping the QTLs linked to this trait.

### **DNA markers**

With the completion of the genome sequence (RiceGAAS) for rice (*Oryza sativa* L.), the focus of rice genomics research has shifted to the comparison of the rice genome with genomes of other species for gene cloning, breeding, and evolutionary studies. Studies were undertaken in *Sorghum bicolor*, a species which diverged from cultivated rice 40–50 million

years ago (Gaut BS, 2002). Hybridisation-based markers (overgos), in conjunction with fingerprint and BAC end sequence data, were used to build sequence ready BAC contigs for two wild *Oryza* species. When rice overgos were aligned to available *S. bicolor* sequence, 29% of the overgos aligned with three or fewer mismatches; of these, 41% gave positive hybridization signals (Hass-Jacobus et al, 2006). Overgo hybridization patterns supported colinearity of loci in regions of sorghum chromosome 3 and rice chromosome 1 and suggested that a possible genomic inversion occurred in this syntenic region in one of the two genomes after the divergence of *S. bicolor* and *O. sativa*.

### **Expression profiling**

Progress in characterization of the transcriptome has been paralleled by identification of differential gene expression in response to biotic and abiotic factors, including greenbug feeding (Park et al., 2006), dehydration, high salinity and ABA (Buchanan et al., 2005), and methyl jasmonate, salicylic acid, and aminocyclopropane carboxylic acid treatments (Salzman et al., 2005).

### **Association genetics**

Much of the value of the sorghum sequence may be realized through better understanding of the levels and patterns of diversity in extant germ plasm, which can contribute both to functional analysis of specific sorghum genes and to deterministic improvement of sorghum for specific needs and environments. Sorghum is well suited to association mapping methods because of its medium-range patterns of linkage disequilibrium (Hamblin et al, 2005) and its self-pollinating mating system.

Extensive *ex situ* sorghum germplasm collections exist within the U.S. National Plant Germplasm System and ICRISAT. Early characterization of complementary association genetics panels developed by a group of US scientists and by Subprogram 1 of the Generation Challenge Program, is in progress. At present, more than 750 SSR alleles and 1402 SNP alleles discovered in 3.3Mb of sequence (Hamblin et al., 2005; Casa et al., 2005) are freely available from the *Comparative Grass Genomics Center* relational database. Extensive studies of sequence variation in sorghum show that haplotype diversity is low, even when nucleotide diversity is high: for regions of average length 671 bp surveyed in 17 accessions, the median number of haplotypes was three and the mode was two (Hamblin et al, 2005). Common sequence variation can therefore be captured in a small sample of accessions.

### **Genetic mapping**

Linkage mapping in sorghum takes advantage of its straightforward diploid genetics, amenability to inbreeding, high levels of DNA polymorphism between *Sorghum* species, and manageable levels of DNA polymorphism within *S. bicolor*. High-density reference maps of one intraspecific *S. bicolor* (Klein et al., 2000; Menz et al., 2002) and one interspecific *S. bicolor* x *S. propinquum* (Bowers et al., 2003) cross provide about 2600 sequence-tagged-sites



(based on low-copy probes that have been sequenced), 2454 AFLPs, and 1375 sequence-scanned based on sequences of genetically anchored BAC clones) loci. These two maps share one common parent (*S. bicolor* “BTx623”) and are essentially collinear (Feltus et al, 2006). Cytological characterization of the individual sorghum chromosomes has provided a generally adopted numbering system (Kim et al, 2005). More than 800 markers mapped in sorghum are derived from other taxa (hence serve as comparative anchors) and additional sorghum markers have been mapped directly in other taxa, or can be plotted based on sequence similarity. Anchoring of the sorghum maps to those of rice, maize (Bowers et al., 2003) sugarcane (Ming et al, 1998), millet, switchgrass (Missaoui et al, 2005), bermuda grass, and others provides for the cross utilization of results to simultaneously advance knowledge of many important crops.

### **Physical mapping**

Sorghum was the first angiosperm for which a BAC library was published (Woo et al, 1994). Estimates of the physical size of the sorghum genome range from 700 Mbp based on Cot analysis (Peterson et al, 2002) to 772Mbp based on flow cytometry. This makes the sorghum genome about 60% larger than that of rice, but only about 1/4 the size of the genomes of maize or human. Peterson et al, (2002) using DNA renaturation kinetic analysis showed the comparative composition of the sorghum genome.

### **Genome sequence**

The shotgun sequencing of a leading US sorghum inbred, BTx623, is now complete, with  $\approx 10.5$  million reads ( $\approx 8X$  coverage) deposited in the NCBI Trace Archive. Early analysis confirms that the sorghum genome sequence will be a suitable substrate for a complete and high-quality annotation. Alignments of the preliminary assembly to sorghum methyl-filtered sequence; sorghum, maize, and sugarcane transcript assemblies; and the Arabidopsis and rice proteomes confirms the base-level accuracy of the assembly and correct local structure of protein-coding loci. Additional resources from reduced-representation sequencing will contribute to the identification of expressed portions of the genome sequence. The sorghum gene space is presently represented by approximately 204 000 expressed sequence tags, many of which have been clustered into  $\approx 22000$  unigenes representing more than 20 diverse libraries from several genotypes (Pratt et al, 2005). About 500 000 methyl-filtered (MF) reads that provide an estimated 1X coverage of the MF-estimated gene space (Bedell et al, 2005) have been assembled into contigs (SAMIs, <http://magi.plantgenomics.iastate.edu>).

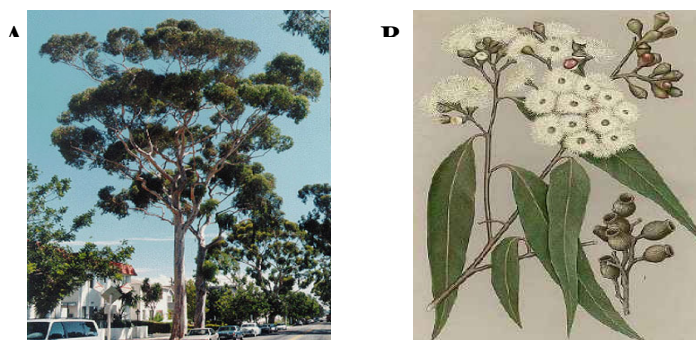
## LIGNOCELLULOSIC FEEDSTOCKS (WOODY & HERBACEOUS)

### *Eucalyptus spp.*

#### Introduction

Native to the Australian continent and its northern neighbours, Eucalyptus is the most widely planted hardwood tree in the world and constitutes one of the world's main sources of biomass. Its versatility and rapid annual growth are still being explored in plantations already estimated at 19 million ha, spread over 37 countries and accounting for 16% of forest plantation areas worldwide (Carbonnier, 2004). India is the largest planter (8 million ha), followed by Brazil (3 million ha) (Junghans et al., 2003), while in Australia and particularly Tasmania, there has also been a move towards growing Eucalyptus in plantations in addition to the 41 million ha of natural Eucalyptus forest (Junghans et al., 2003). Amongst 700 Eucalyptus species, while *E. grandis* is the most widely cultivated species in subtropical and warm temperate regions, *E. camaldulensis* is the most common species in arid and semi-arid lands and *E. globulus* is the main species in temperate climates free of severe frosts. Elite clones are mainly used in Brazil by the cellulose and paper industry because of wood quality and high volume yield. Mature trees also provide strong and durable timber which is the major use of Eucalyptus, along with fuel supply, in countries such as India. The name *eucalyptus* comes from the Greek: *ευκάλυπτος*, *eukályptos*, meaning "well covered", or "beautiful bark" according to linguist Ahmed Seddik.

*Eucalyptus* has attracted attention from global development researchers and environmentalists. It is a fast-growing source of wood, its oil can be used for cleaning and functions as a natural insecticide, and it is sometimes used to drain swamps and thereby reduce the risk of malaria. Outside their natural ranges, eucalypts are lauded for their beneficial economic impact on poor populations and derided for being invasive water-suckers, leading to controversy over their total impact.



**Figure 1.** A) A mature *Eucalyptus* tree; B) Flowers and seeds of *Eucalyptus maculata* Hook

## **Biotechnology approaches**

### **Genome research**

Eucalypts are diploid plants with a haploid chromosome number of 11. The size of the genome has been estimated for several eucalypt species and their hybrids to range from 370 to 700 million Mbps.

The nuclear genome of eucalypts has been represented by maps generated through linkage analysis of DNA based polymorphic markers (Shepherd & Jones, 2005). These markers include microsatellites or simple sequence repeats (SSRs), RAPDs, AFLPs and RFLPs ((Thamarus et al., 2002). Some of these markers are dominant, such as RAPDs and AFLPs. Microsatellites and RFLPs on the other hand are codominant markers and are generally more informative as they are multiallelic. Although dominant markers are relatively cheap they have limited transferability, whereas codominant markers have a much broader transferability and are potentially more informative in crosses with up to four alleles segregating such as outcross F2 and pseudo-testcrosses (Bundock et al., 2000).

Organelle genome structure and inheritance has also been investigated in eucalypts. Studies have found the chloroplast to be maternally inherited in eucalypts (McKinnon et al., 2001b). By comparing the *E. globulus* chloroplast genome sequence with that of other sequenced species including *Nicotiana tabacum* and *Oenothera elata*, very high homology was identified in the coding regions between these species and high divergence in the intragenic regions, which were also a source of microsatellites. Eucalypt mitochondria have been less well characterized. It has been reported that the mitochondria are maternally inherited in *E. globules*. Rengel et al. (2009) have reported the sequencing, assembly and annotation of approximately 10,000 ESTs derived from a normalized full-length secondary xylem cDNA library as well as subtractive libraries. The EST assembly generated a set of 3,857 wood-related unigenes including 2,461 contigs (Cg) and 1,396 singletons (Sg) that were named 'EUCAWOOD' creating a valuable resource for functional genomics studies of wood formation and molecular breeding in this economically important genus. About 65% of the EUCAWOOD sequences produced matches with poplar, grapevine, *Arabidopsis* and rice protein sequence databases. The number of sequences has significantly increased (Costa da Cruz et al., 2008; Novaes et al., 2008; Qiu et al., 2008) but is still low in comparison to other forest tree species such as poplar or pine. The major part of this new data set is composed of short sequences whose number is expected to increase dramatically in the future thanks to the development of the high throughput '454' technology (Novaes et al., 2008).

Species	Modified trait	Gene(s) altered	References
<i>E. camaldulensis</i>	Cellulose	cbd, cell1	Shani et al., 2003
<i>E. camaldulensis</i>	Lignin	C4H, CAD, Ntlm1	Chen et al., 2001; Kawaoka et al., 2003; Valerio et al., 2003
<i>E. camaldulensis</i>	Stress resistance	DREB1A	Hibino et al., 2002; Kondo et al., 2002 ; 2003
<i>E. camaldulensis</i>	Salt stress tolerance	cod A	Yamada-Watanabe et al., 2003
<i>E. camaldulensis</i>	Insect/herbicide resistance	cry3A, bar	Harcourt et al., 2000
<i>E. grandis</i>	Cellulose	cbd, cell1	Shani et al., 2003
<i>E. grandis</i> · <i>E. urophylla</i>	Lignin	CAD	Tournier et al., 2003
<i>E. urophylla</i>	Bacterial wilt resistance	cecropin D	Shao et al., 2002

**Figure 2:** Research involving genetic modification in *Eucalyptus* species (Poke et al., 2005)

### Quantitative Trait Locus Mapping

The construction of a comprehensive microsatellite-based linkage map for commercial species of *Eucalyptus* is now well advanced (Brondani and Grattapaglia, 2002). In the context of Genolyptus project (Grattapaglia, 2004) a target was established to develop and map 1,000 microsatellite markers. Sources for this large number of microsatellites are now becoming available, not only from the enriched library approach, but also from a shotgun genomic library, a large set of ESTs, and from BAC end sequences. The availability of transportable, multiallelic, PCR-based co-dominant microsatellite loci provides a fundamental tool to carry out linkage and quantitative trait locus (QTL) analysis in eucalypts and allows researchers to move from phenotypes to target genomic regions controlling traits of interest.

Transcript abundance, measured for 2,608 genes in the differentiating xylem of a 91 (*E. grandis* X *E. globulus*) X *E. grandis* backcross progeny was correlated with diameter variation, revealing coordinated down-regulation of genes encoding enzymes of the lignin biosynthesis and associated methylation pathways in fast growing individuals (Kirst et al., 2004). Quantitative trait locus (QTL) analysis of transcript levels of lignin-related genes showed that their mRNA abundance is regulated by two genetic loci, demonstrating coordinated genetic control over lignin biosynthesis.

### Association mapping

Genetic mapping became accessible to several forest tree species in the beginning of the 90's based on the combination of the speedy and inexpensive generation of dominant RAPD and AFLP markers and the pseudo-testcross strategy in two-generation pedigrees or the use of the haploid genetics of conifers. Concomitant to this development, linkage maps of co-dominant markers led to the construction of integrated RFLP maps for a few species (Devey et al., 1994) and the possibility of comparative mapping (Komulainen et al., 2003; Krutovsky et al., 2004). However it soon became clear that true advancements in QTL validation across pedigrees and eventually marker assisted selection in forest trees, would strongly depend on the

availability of higher throughput, higher polymorphism typing systems such as microsatellites, organized in dense genetic maps (Brondani et al., 1998; Grattapaglia, 2000). In the last few years a number of studies reported genetic maps for forest trees built with combinations of several hundred RAPD and AFLP markers together with some tens of EST, genes and microsatellites (Brondani et al., 2002; Achere et al., 2004). Linkage maps with around one hundred microsatellites were reported for *Pinus taeda* (Zhou et al., 2003) and *Populus* (Yin et al., 2004). However to allow a more precise comparison of QTL position and validation of putative QTL across pedigrees larger sets of microsatellites are clearly necessary.

Using marker-based methods, significant narrow-sense heritability of foliar defense chemicals in a natural population of *Eucalyptus melliodora* was found (Andrew et al., 2005).

With the rapid advancement of genome projects generating a large amount of sequence information and single nucleotide polymorphism (SNP) (one-letter variations in the DNA sequence that contribute to differences among individuals) data, plant genomics has experienced a growing interest in an alternative approach for the identification of genes underlying quantitative traits. Some laboratories have started association mapping work for wood traits, both in Pines (Brown et al., 2001) and in *Eucalyptus* (Thumma et al., 2005), by sampling trees in the wild or from breeding programs that display contrasting phenotypes for wood quality traits. Külheim et al., (2009) discovered 8,631 SNPs across the species *Eucalyptus globulus*, *E. nitens*, *E. camaldulensis* and *E. loxophleba*.

Another marker-based technique is the one that uses microsatellites for the detection of differences and mapping of *Eucalyptus* species genetic backgrounds. One hundred thirty seven autosomal microsatellite markers have been published to date for species of *Eucalyptus* and 70 from *E. grandis* and *E. urophylla* (Brondani et al., 2002). Recently, a set of 35 chloroplast DNA microsatellites were developed based on the full cp-DNA sequence of *E. globules*. Bordani et al. (2006) reported the construction of a consensus genetic linkage map covering all 11 linkage groups of *Eucalyptus* including a total 234 mapped loci making it, according to their knowledge, the most complete genetic map of *Eucalyptus* and of a forest tree to date based exclusively on interspecific transferable microsatellites.

### **EST programmes**

Similar to the efforts in poplar, sequencing projects often form part of large genomics programs undertaken by national or international consortia. The biggest on-going joint public and private programs were founded in Brazil, demonstrating the economic importance of *Eucalyptus* at the state level in this country. The Genolyptus project (<http://genolyptus.ucb.br/genolyptus-english.jsp>), which includes thirteen companies, seven universities and Embrapa (Brazilian Enterprise for Agricultural Research) is targeting wood quality and disease resistance. Divided into several topics (Grattapaglia, 2004), the research program aims to translate genomics knowledge into improved tree breeding technologies. In

2004, the sequencing project reached about 150,000 ESTs for *E. grandis* (50%), *E. globulus* (16%), *E. urophylla* (10%) and *E. pellita* (10%), the remaining 14% for six other species.

The second Brazilian consortium, FOREST (<http://est.cbmeg.unicamp.br/pgl/research/forests.html>), is made up of twenty laboratories from San Paulo and four companies. The isolation of 124,000 sequences (12 cDNA libraries) was obtained mainly from *E. grandis*, representing different organs under different growth conditions and related to environmental stress tolerance (Furtado et al., 2004). In addition, OJI Paper has 60,000 ESTs.

Another independent French public genome sequencing program (26,000 ESTs from *E. gunnii*) was also named FOREST (<http://www.genoscope.cns.fr>). These Eucalyptus sequences were isolated in the framework of the Toulouse University - CNRS research program, focusing on the regulation of lignin biosynthesis ([http://www.smcv.ups-tlse.fr/root/equipes/regulation/equipe\\_en.php](http://www.smcv.ups-tlse.fr/root/equipes/regulation/equipe_en.php)) and frost tolerance ([http://www.smcv.ups-tlse.fr/root/equipes/stressfroid/equipe\\_en.php](http://www.smcv.ups-tlse.fr/root/equipes/stressfroid/equipe_en.php)). The sequences are being released on GenBank. In Australia, the CSIRO forestry research programs (<http://www.ffp.csiro.au/tigr/molecular/>) led to the isolation of EST collections from *E. grandis* and *E. grandis* X *E. nitens* hybrids. A current research project on wood quality is based on a 5000 cDNA library from Eucalyptus xylem. Moreover, the same research groups are also interested in flowering and cold tolerance (Fullard and Moran, 2003). At the University of Melbourne, 93 sequences of cambial genes from *E. globulus* have been posted in GenBank (accession no AW191301–AW191393) and, from them, 43 could be annotated (Bossinger and Leitch, 2000).

### **Tissue Culture of Eucalyptus**

Conventionally, eucalyptus is propagated through seeds. However, due to segregation of genes, the seed-raised population is highly heterogeneous. Most Eucalyptus can be propagated vegetatively using traditional stem cutting techniques. However, in vitro micropropagation and rooting, followed by transfer to soil, is becoming increasingly popular because of the prospects of rapid genetic gain. These micropropagated plants are not necessarily used in the establishment of plantations, but are useful source material for the development of clonal hedges and hydroponic systems as sources of cuttings. Micropropagation by microcuttings is commonly carried out on *E. camaldulensis*, *E. globulus*, *E. grandis*, *E. nitens*, *E. tereticornis*, *E. urophylla* and *E. gunnii*. In addition to these conventional multiplication procedures, the production of somatic embryos, which renders micropropagation highly cost effective, would be a powerful tool for the establishment of plantations of elite Eucalyptus genotypes (Bandyopadhyay et al., 1999; Bandyopadhyay and Hamill, 2000). Dhawan and Saxena (2004) have been successful in multiplying three species of eucalyptus i.e., *E. tereticornis*, *E. camaldulensis* and *E. citriodora*.

### **Hybridization**

Molecular genetics can play a major role in understanding hybrid inviability and identifying compatible individuals and in implementing advanced generation hybridization. For example, DNA markers showed that high levels of segregation distortion occur in both a selfed *E. gunnii* x *globulus* F2 (Vaillancourt et al., 1995) and an *E. grandis* x (*E. grandis* x *globulus*) backcross (Myburg et al., 2000).

There is also a clear role for marker-assisted selection (MAS) in advanced generation hybridization strategies (Griffin et al., 2000). Marker assisted backcrossing has great potential to speed the introgression of favourable genes into the recurrent species (Myburg et al., 2000). It is being proposed to aid with the introgression of salt tolerant genes into *E. grandis* and *E. globulus* through hybridization with salt-tolerant selections of *E. camaldulensis* in Australia (Dale et al., 2000).

## ***Populus spp.***

### **Introduction**

*Populus* is a genus of 25–35 species of deciduous flowering plants in the family Salicaceae, native to most of the Northern Hemisphere. English names variously applied to different species include poplar, aspen, and cottonwood. The genus has a large genetic diversity, and can grow from anywhere between 15–50 m tall, with trunks of up to 2.5 m diameter. In addition to their value for wood products, members of the genus *Populus* (poplars) provide a range of ecological services, including carbon sequestration, bioremediation, nutrient cycling, biofiltration and diverse habitats. They are also widely used model organisms for tree molecular biology and biotechnology. The sequencing of the poplar genome to an approximately 6X depth adds to a long list of important attributes for research. These include facile transformation, vegetative propagation, rapid growth, modest genome size and extensive expressed sequence tags.

### **Biotechnology approaches**

#### **Genome sequencing**

The publication of the draft sequence of *Populus* (Tuskan et al., 2006) based on the genotype Nisqually-1 creates a new resource for bioenergy applications in that it contains all the genes to build a tree. Breaking the code requires more work than simply obtaining the DNA sequence itself – the process of decoding the genome sequence is called annotation (Stein, 2001). The challenge in poplar is to identify the various alleles that control trait variation within the species or interspecific pedigree of interest, since each gene may have many alleles within each species.

Genes that condition plant responses to the hormones auxin and cytokinin, known to have dramatic effects on tree growth, development and architecture, have also been annotated

(Kalluri et al., 2007; Ramirez-Carvajal et al., 2008). There is also the annotation of gene families that belong to mitogen-activated kinase family (Nicole et al., 2006), to the heat shock protein family (Waters et al., 2008) and to the LIM protein family (Arnaud et al., 2007).

### **QTLs**

Most bioenergy-related traits are quantitative, meaning that they are typically either oligogenic (few to several loci regulate variation in the trait) or polygenic (many loci are involved) with a measurable effect of the environment on trait expression. Quantitative trait loci (QTL) associated with variation in these traits can be identified using QTL mapping. When QTL are identified in a well-designed study, it reinforces that the trait is heritable, and simultaneously identifies a genomic region that contains the gene(s) involved.

QTL for stem height, stem circumference, stem volume, number of sylleptic branches and total aboveground biomass were recently identified (Rae et al., 2008) and these data complement earlier studies that identified QTL for aboveground, belowground, leaf, stem and coarse root biomass, as well as the proportion of biomass allocated aboveground to leaves and to stems, and allocated belowground, and belowground specifically to coarse roots, and into fine roots (Wullschleger et al., 2005). In a long-lived species such as poplar, it is also essential to understand how biomass production changes with maturity (or in the case of SRC, the individual stools and the entire stand) with understanding being required at the genetic and physiological/morphological level. A number of studies have reported QTL in the population used for this study at a single time point, for single stem plants usually during early phases of growth. More recently, a QTL mapping identified regions of genetic control for biomass yield in poplar (Rae et al., 2009).

### **EST programmes**

Poplars deploy an array of combined defense strategies against herbivores that can be grouped as chemical and physical defenses, direct and indirect defenses, constitutive and induced defenses, as well as local and systemic defenses. Several recent studies have been conducted on the molecular mechanisms underlying inducible defenses against herbivores in poplar (Major and Constabel, 2006; Miranda et al., 2007). Another study was the gene expression analysis in autumn leaves of poplar, in which Bhalerao et al (2003) preparing cDNA libraries and obtaining ESTs, identified genes that are involved only in the leaf senescence.

The poplar EST database from Umea Plant Science Centre (Umea, Sweden) was used by Ferreira et al. (2006) in order to contribute to the knowledge of the molecular mechanisms underlying Euphrates poplar tolerance/resistance to high temperatures. In September 2004, opened to the public a *Populus* expressed sequence tag (EST) database (POPULUS DB) which was created from 19 cDNA libraries each originating from different *Populus* tree tissues (Sterky et al., 2004). The database consists of 102,019 ESTs, assembled into a unigene set of 11,885 clusters and 12,759 non-clustered singletons corresponding altogether to 24,644 unique



sequences or transcripts (POPULUSDB). Moreau et al (2004) used this dataset for in silico transcript profiling of a particular process in the woody tissues of the *Populus* stem: the programmed death of xylem fibers.

As part of the poplar genome sequencing project and the development of genomic resources for poplar, Ralph et al (2008), generated a full-length (FL)-cDNA collection using RNA from xylem, phloem and cambium, and green shoot tips and leaves from the *P. trichocarpa* Nisqually-1 genotype, as well as insect-attacked leaves of the *P. trichocarpa* X *P. deltoides* hybrid.

ForestTreeDB is intended as a resource that centralizes large-scale expressed sequence tag (EST) sequencing results from several tree species (<http://foresttree.org/ftdb>) (Pavy et al, 2006). It currently encompasses 344878 quality sequences from 68 libraries from diverse organs of conifer and hybrid poplar trees, and aims to be continuously enriched. In an earlier study, around 14,000 expressed sequence tags (ESTs) representing genes involved in abiotic stress responses from several normalized and subtracted cDNA libraries produced from control, stress- exposed ,and desert-grown *P.euphratica* trees were sequenced (Brosche et al., 2005). A microarray with a unigene set of 6,340 ESTs enriched in stress-related genes was constructed (Brosche et al., 2005) and used in the characterization of the transcriptional responses to gradual soil water depletion (Bogeat-Triboulot et al., 2007).

### **Microarrays**

Transcript profiling using microarrays has made possible to test the potential involvement of thousands of genes in a biological process, thus providing valuable information for the selection of target genes. This technology has already generated important data on gene expression profiles during the transdifferentiation of mesophyll cells into xylem cells in *Zinnia elegans* and identified candidate genes involved in xylem formation in hybrid aspen (*Populus tremula* x *tremuloides*) (Hertzberg et al., 2001). Schrader et al. (2004) have reported a high-resolution transcript profile across the cambial zone of aspen (*Populus tremula*) for more than 13,000 genes. Ninety-five publicly available DNA microarray datasets were obtained from the Gene Expression Omnibus (GEO) database (<http://www.ncbi.nlm.nih.gov/geo/>) by Ogata et al., (2009). They performed a co-expression network analysis using the datasets and extracted “co-expression modules,” comprising co-expressed genes, which are tightly interconnected to each other. Information on the experiments contributing to gene-to-gene connections in co-expression modules was associated with the modules. Their database is available at <http://webs2.kazusa.or.jp/kagiana/cop/>.

### **Transformation system**

A very widely used transformation method to produce transgenic poplar plants is that of the *Agrobacterium tumefaciens* transformation method (Arisi et al., 1997; Noctor et al., 1998). New advances have been achieved in plastid transformation. In most angiosperm plant

species, plastid genes are maternally inherited, and therefore, transgenes in these plastids are not disseminated by pollen. This makes plastid transformation a valuable tool for the creation and cultivation of genetically modified plants that are biologically contained, thus posing lower environmental risks (Daniell, 2007). Okumura et al. (2006) reported an efficient chloroplast transformation method by bombarding poplar leaves with 0.6- $\mu$ m gold particles. However, the most recent transformation technique that has been reported is the one of Liu et al (2009) in which they succeeded the transformation of whole poplar plants by exposing them hydroponically to 3, 3', 4, 4'-tetrachlorobiphenyl (CB77).

### **Characteristics of interest**

Over the past few years, there have been several reports regarding the study of poplar's lignin content and its metabolic pathways (Poke et al., 2006; Leple et al., 2007).

Many characteristics of growth have also been studied in poplar. Transgenic manipulation of poplar growth was accomplished by enhancing biosynthesis of the phytohormone gibberellin through overexpression of the gene encoding GA 20-oxidase (Eriksson et al., 2000) and more recently, genes were identified that play crucial role in the perception of light and the regulation of the circadian clock (Loivamki et al, 2007).

*Populus* species (cottonwoods, poplars, and aspens, hereafter referred to collectively as poplar) are often ecological foundation species and include the most widely distributed trees in the Northern Hemisphere. The phenolic metabolites produced by poplar are thought to be important determinants of community structure and ecosystem dynamics (Whitham et al., 2006). Poplar leaves typically accumulate several classes of phenolic metabolites, including the salicylate-derived phenolic glycosides (PGs), flavonoids such as flavonol glycosides, anthocyanins, and proanthocyanidins (PAs; or condensed tannins), and numerous small phenolic acids and their esters (Lindroth and Hwang, 1996). In the literature, there are many reports available on the study of flavonoid and PA biosynthetic pathways genes (Mellway et al., 2009; Wilkins et al., 2009).

## ***Cynara cardunculus* L.**

### **Introduction**

*Cynara cardunculus* L. is a diploid ( $2n=34$ ) out crossing perennial species native to the Mediterranean basin. It belongs to the Asteraceae family and includes the globe artichoke (var. *scolymus* L.), the cultivated cardoon (var. *altilis* DC), and the wild cardoon (var. *sylvestris* (Lamk) Fiori). Molecular, cytogenetic and isozyme evidence suggests that *Cynara cardunculus* var. *sylvestris* is the wild ancestor of the globe artichoke and cultivated cardoon (Rottenberg A and Zohary, 1996; Raccuia et al., 2004). While the former is vegetatively propagated, the latter is propagated by seeds. The existence of this complex primary genepool, containing the wild progenitor and the two crops showing different reproductive strategies, is unique among crop

species, making the study of genepools and germplasm differentiation particularly interesting for the utilization of *Cynara* genetic resources (Pagnota and Noorani, 2010). The crop remains of regional importance in Spain, Italy, Greece and the south of France, where it is used in traditional dishes. In spite of its old origin and its good flavour the cardoon has never become a widespread crop. For instance in Spain, that is one of the countries that most cultivated cardoon, has only 1000 ha (96% irrigated lands) for this crop. Named varieties are difficult to find outside the Mediterranean region. Wild and cultivated forms of *C. cardunculus* are allogamous and perennial.



**Figure 1.** *Cynara cardunculus* L.

## **Biotechnology approaches**

### **Genome mapping**

The genome research of *Cynara cardunculus* L, unlike other species belonging to Asteraceae (Compositae) family (i.e. sunflower, lettuce and chicory), is far behind. The species is highly heterozygous and suffers marked inbreeding depression when forced to self-fertilize.

Artichoke breeding programs have traditionally aimed at improving earliness, head yield and quality, resistance to disease (such as *Verticillium*) and nematodes (Arce et al., 2004; Miguel et al., 2004; Esteva et al., 2004). Breeding programmes have been based on intracloonal selection (Pècaut 1983; Mauromicale and Copani 1989) or hybridization among varietal groups followed by selection (Baznisky and Zohary, 1994). Therefore, efforts have been made in cynara genome research, especially development of genomic resources and tools for basic and applied genetics, genomics, and breeding research. These resources and tools include different types of DNA markers such as randomly amplified polymorphic DNA (RAPD) (Lanteri et al., 2001; Sonnante et al., 2002), amplified fragment length polymorphism (AFLP) (Lanteri et al., 2004b, Portis et al., 2005a; 2005b) and simple sequence repeat (SSR) or microsatellites (Acquadro et al., 2005a; 2005b, 2009).

Recently, large-scale sequencing work in *Cynara cardunuculus* has been generating about 33M of pair-end 75bp sequences corresponding to 2,500 Mbp (2.3X genome coverage of artichoke) (Sonnante et al., 2011) and a collection of 36,321 ESTs, generated from the 'Green Globe' variety of *C. cardunculus* var. *scolymus*, as part of the output of the Compositae Genome Project ("<http://compgenomics.ucdavis.edu>") (Scaglione et al., 2009). The

cataloguing of these sequences in public databases is providing useful information to develop markers with high resolving power, such as SNPs thus initiating an era of *in silico* cynara marker discovery.

### **Molecular markers**

In order to understand the function of specific genes and their role in metabolic pathways, as also to identify the key steps in their coregulation mechanisms, several approaches have been exploited. The identification of the genetic basis of metabolite variation in *A. thaliana* has been pioneered by Keurentjes et al. (2006), by applying quantitative trait loci (QTL) analyses on a large metabolomics data set. However, the *C. cardunculus* genome is still poorly mapped. In order to move to a crossing strategy for breeding, a greater knowledge of globe artichoke genome will be essential.

There has been reported the isolation and characterization of a gene involved in involved in both chlorogenic acid and lignin biosynthesis (Moglia et al., 2009) and an enzyme involved in the phenylpropanoid pathway (De Paolis et al., 2008). Moreover, gene sequences encoding hydroxycinnamoyltransferase (HCT and HQT), involved in the synthesis of chlorogenic acid, have been recently identified, characterized and incorporated within the developing globe artichoke linkage maps (Comino et al., 2009).

### **Comperative proteomics**

Although molecular markers have been developed and applied to produce a genetic map the *Cynara cardunculus* genome remains poorly researched. The genome is fixed in time, but the proteome is very plastic, depending on tissue type, developmental stage and age, and is also strongly modulated by the environment. The differential response of related proteomes to the same set of biotic and abiotic factors allows the genetic mapping of expressed genes (Thiellement et al., 2002). A proteomic analysis compares gene products involved across the full range of physiological processes, and illustrates the dynamic nature of cell/tissue processes (Rajjou et al., 2006). The first leaf proteome analysis for globe artichoke has been performed (Acquadro et al., 2009) and mass spectrometry- (MS-) was used for the identification of selected protein spots. Comparison of global protein expression profiles of green and etiolated fleshy stalks of cultivated cardoon indicated that, during etiolation, the differentially expressed proteins were involved mainly in starch metabolism and stress response (Guarino et al., 2010).

### **Tissue culture**

Generally, globe artichoke is propagated vegetatively by offshoots, stumps or dried shoots harvested from commercial fields at the end of the production cycle; however, the potential for the spread of pest (nematodes, fungi and viruses) using the current propagation technique is very high leading to significant economic losses. It has been demonstrated that plants obtained with the meristem tip culture technique shown improved field performance with

respect to both qualitative and quantitative traits, and this can compensate for the higher cost of the planting material (Saccardo et al., 2007).

The availability of an efficient protocol for the *in vitro* production of haploid plants and subsequent diploidisation would greatly speed the development of the homozygous material needed for F1 hybrid breeding (Lanteri and Portis, 2008). The first reported attempts to culture anthers from five Italian cultivars resulted only in the production of callus (Motzo and Deidda, 1993). Although microspores can now be reproducibly cultured, development beyond the second division has not yet been attained, presumably because of non-optimal culture conditions (Stamigna et al., 2004). Haploid production via gynogenesis has been also been unsuccessful although *in situ* gynogenesis using fertilisation with irradiated pollen has been reported by the INRA station (Lanteri and Portis, 2008). However, this method is at present not sufficiently reproducible for general use (Stamigna et al., 2004).

## FIBER CROPS

### *Cannabis sativa* L.

#### Introduction

*Cannabis sativa* is an annual plant in the Cannabaceae family. It's one of the oldest known domesticated plants and today is cultivated throughout the world for psychoactive cannabinoids, durable fiber, and nutritious seed. Different parts of the plant have different uses, and different varieties are cultivated in different ways and harvested at different times, depending on the purpose for which it is grown. Cannabis can be separated into psychoactive and nonpsychoactive cultivars according to the ratio of D9-tetrahydrocannabinol (THC,) the primary psychoactive agent, and cannabidiol (CBD) (Hillig and Mahlberg, 2004).



**Figure 1:** *Cannabis sativa* L. Plant parts and seeds.

Hemp is a dioecious, herbaceous annual plant with a four to six month growing season. Dioecy, by definition, means that pistillate (female) and staminate (male) flowers are presented on separate plants. Marijuana can be propagated in two ways: by seed or by cloning. Seeds are a result of sexual reproduction between a pistillate and staminate plant and produce new individuals with recombinant genotypes. Isolated female plants will produce prolific floral buds with a high THC content. Cloning is a form of asexual reproduction that allows for preservation of the genotype due to lack of meiotic recombination. This form of propagation is desirable to the grower because it perpetuates the unique characteristics of the parent plant. It also generates a population of nearly identical, all-pistillate, fast-growing and evenly maturing Cannabis plants. To propagate marijuana by cloning, a cutting is removed from the parent plant and induced to form a new root system. Root systems typically develop in three to six weeks and the clones are then ready to be transplanted into larger containers. Plant development can be accelerated by supplying excess nutrients, carbon dioxide and light. With a sudden shift from twenty-four hour daylight to a twelve hour light regime to mimic autumn conditions, marijuana plants can be forced to flower before they are eight weeks of age.

## **Biotechnology approaches**

### **Inheritance of traits**

The class of secondary products unique to the dioecious species *Cannabis sativa L.* (hemp) is the terpenophenolic substances known as cannabinoids, which accumulate mainly in the glandular trichomes of the plant (Hammond and Mahleberg, 1977). Over 60 cannabinoids are known (de Zeeuw et al., 1972a), the most abundant being cannabidiol (CBD) and D-9-tetrahydrocannabinol (THC). Small and Beckstead (1973) were the first to systematically survey a wide number of Cannabis accessions for variability in cannabinoid composition. De Meijer et al (1992), in a survey of large Cannabis collection, also found that plants belonging to the same population often show distinct CBD/THC ratios.

### **DNA markers**

Today, the concept of Cannabis as a monotypic genus is widely accepted; taxonomical, morphological and biometrical studies confirm the continuity of its gene pool despite the extremely high variation found within and between populations. In the last few years, the existence of just a single species within the genus has been confirmed by molecular marker studies that show a limited segregation of the different groups within the genus Cannabis and an extremely high degree of polymorphism, estimated to be of the same magnitude within and between populations (Faeti et al., 1996; Forapani et al., 2001). Within the dioecious populations, the presence of a high number of male-specific markers, presumably associated with the Y chromosome, was found by RAPD and amplified fragment length polymorphism (AFLPs) analysis (Mandolino et al., 2004; Flachowsky et al., 2001). Alghanim and Almirall

(2003) developed 11 microsatellite markers that found to be useful for DNA typing and for assessing genetic relatedness in *Cannabis*.

### **Improvement of bio-components of interest**

Hemp is thought to be likely the first plant cultivated by mankind for its textile use (Lu and Clarke, 1995). The methods today for modification of hemp fibers, in order to make them finer, cleaner, softer and more suitable for processing on machines of higher efficiency than traditional hemp machines, are chemical, chemomechanical and mechanical methods. One recently advanced method is the modification of hemp fibers with sodium hydroxide solutions under different conditions, in order to partially extract lignin, pectins and hemicelluloses, and separate the fiber bundles (Kostic et al, 2008). The quality of modified hemp fibers can be characterised by determining their chemical composition, fineness, mechanical and sorption properties.

### **Distinguishing Hemp from Marijuana**

Currently available methods of analysis for THC in Cannabis leaf material are high-performance liquid chromatography (HPLC) with UV detection (Rustichelli et al., 1998), gas chromatography with flame-ionization detection (GC-FID) (de Meijer et al., 1992), and a screening method based on HPLC. These methods are used for purposes like the detection of higher than the maximum allowed concentration of the psychoactive cannabinoid,  $\Delta^9$ -tetrahydrocannabinol, in industrial fiber hemp. Additionally, Hewavitharana et al. (2005) developed a new method based on mass spectrometry (MS) by which the total THC concentration can be determined accurately leaving outside closely or co-eluting compounds from the sample.

## ***Hibiscus cannabinus* L.**

### **Introduction**

Kenaf (*Hibiscus cannabinus* L., *Malvaceae*) is a warm season annual fiber crop closely related to cotton (*Gossypium hirsutum* L., *Malvaceae*) and okra (*Abelmoschus esculentus* L., *Malvaceae*) that can be successfully produced in a large portion of the United States, particularly in the southern states. As the commercial use of kenaf continues to diversify from its historical role as a cordage crop (rope, twine, and sackcloth) to its various new applications including paper products, building materials, absorbents, and livestock feed, choices within the decision matrix will continue to increase and involve issues ranging from basic agricultural production methods to marketing of kenaf products. These management decisions will require an understanding of the many different facets of kenaf production as a fiber, feed, and seed crop.



**Figure 2.** Leaves and plants of kenaf cultivars ‘Everglades 41’ (top and left) and ‘Tainung #2’ (bottom and right).

### **Biotechnology approaches Transformation techniques**

A very efficient method for transformation of kenaf plants using the *Agrobacterium tumefaciens* has been developed (Banks et al., 1993). More recently, the influence of *Agrobacterium* strain, temperature, host tissue wounding, acetosyringone, *virG/virE* genes and host cell division on T-DNA expression in the kenaf shoot apex were investigated (Srivatanakul et al., 2001). Besides the conventional method of *Agrobacterium* transformation using callus or cells in tissue culture, efforts have been made for *in planta* transformation of kenaf (Kojima et al., 2004). Reports on protocols for isolation, electrofusion and culture of kenaf protoplasts as an initial step in plant improvement strategies have been made (Reichert and Liu, 1996). More recently, Liang et al. (2002) developed a protoplast isolation protocol for kenaf leaf tissue from potted plants in order to study replication of *Hibiscus* chlorotic ringspot virus (HCRSV). Another approach used for the transient transformation of kenaf leaves is the biolistic method (Sanford et al., 1993; Liang et al., 2002).

### **Components of interest**

Lignin and cellulose are the high abundant biopolymers in plant cell wall. In the recent years, the demand of vegetable fibers has increased steadily, due to their good mechanical and biodegradability properties and due to the increase of biocomposite material production. Kenaf (*Hibiscus cannabinus* L.) is an important herbaceous plant cultivated mainly as source of vegetable fibers. In particular, lignin fibers are mainly present in the stem cortical external part (bark); the internal part (core) furnishes light and absorbent wood due to its high content of cellulose. Research groups have been focused on the study of major genes involved in the lignin and cellulose biosynthesis pathways (Ruotolo et al., 2007). In addition to the studies



made for kenaf cell wal, the chemical composition of the essential oil of kenaf (*Hibiscus cannabinus*) has also been examined (Kobaisy et al., 2001).

## ***Linum usitatissimum* L.**

### **Introduction**

Flax (*Linum usitatissimum*), also known as linseed, is a member of the genus *Linum* in the family *Linaceae*. It is native to the region extending from the eastern Mediterranean to India and was probably first domesticated in the Fertile Crescent. Flax was extensively cultivated in ancient Ethiopia and ancient Egypt. In a prehistoric cave in the Republic of Georgia dyed flax fibers have been found that date to 34,000BC. *L. usitatissimum* is an economically important crop that is grown either for its fiber (fiber flax) or for its oil (oilseed flax). Both flax and linseed are specialised developments of a single species, which originates from the Mediterranean and Southwest Asian regions as mentioned above.



**Figure 1.** Organs of the flax plant

The terms flax and linseed are often used interchangeably, in North America ‘flaxseed’ describes flax of a human edible form, and ‘linseed’ describes flax used for industrial oils. For a useful description of the composition of flaxseed, see [http://www.flaxcouncil.ca/FlaxPrimer\\_Chptr1.pdf](http://www.flaxcouncil.ca/FlaxPrimer_Chptr1.pdf).

## **Biotechnology approaches**

### **Tissue and organ culture**

The first report of the species capacity to initiate buds from decapitated hypocotyl sections was made by Link and Eggera (1946). Interestingly, an *in vitro* system was employed to study a pathogen as early as 1957 when the production of aerial-mycelium and uredospores by *Melampsora lini* (Pers) H. Lev. on flax leaves in tissue culture was reported by Turel and Ledingham (1957). Work on improving and optimizing culture medium for this species was reported by Ibrahim (1971) and this work was followed up by a study on biochemical differentiation and phenolic compounds from flax tissue cultures by Liau and Ibrahim (1973). Tissue culture, protoplasts and morphogenes have widened the scope of applications in this species (Gamborg and Shyluk, 1976). This species can be induced to regenerate shoots and roots readily, and though some considerable genotypic variation has been observed, this species can be classed as largely amenable to *in vitro* regeneration.

Flax embryos were one of the first embryos cultivated *in vitro*. Later, the experiments of Erdelska (Erdelska et al., 1973) and Pretova (1986) described flax embryo development *in vitro* from the globular stage onwards.

Flax was shown to be responsive in explant culture systems to a wide range of growth regulators including thidiazuron (Bretagne et al., 1994; Jain and Rashid, 2001).

A further elucidation of the genetic basis of *in vitro* regeneration responses was reported by Bonell and Lassaga (2002). The plasticity of the response of this species has also enabled studies on the derivation of salt-tolerant lines from *in vitro* cultures (McHughen and Swartz, 1984) on differential stress tolerance from somaclonal variants (O'connor et al., 1991) and on inducing albino mutations as a tool for genetic analysis and cell biology (Bretagne-Sagnard et al., 1996). An assessment of induction of somaclonal variation was also performed in 1992. A supplementary study on hydroxyl radical formation during *in vitro* morphogenesis of flax and the effects of free radical chemistry modification on morphogenic and embryogenic response has also been reported (Obert et al., 2004b).

Somatic embryos of flax were first derived from immature zygotic embryos (Pretova and Williams, 1986). Somatic embryogenesis has been further investigated in flax, with a fascinating application of somatic embryogenesis described by Ling and Binding (1992) when this mode of regeneration was described as a means of regeneration from protoplasts. An initial report by Cunha and Ferreira (1996) was followed by a more detailed study by the same authors (Cunha and Fernandes-Ferreira, 1999). In addition, the free sterol content variation during the process of somatic embryogenesis was reported (Cunha and Ferreira, 1997) and this work was extended to the determination of the esterified acids content (Cunha and Ferreira, 2003). Somatic embryos of flax were induced by an indirect method from callus by Tejavathi et al. (2000), but growth progression of the shoot apex was not achieved. Another report was made

by Dedicova et al. (2000) on the regeneration of shoots and embryo-like structures (ELS) from hypocotyl segments. Progress on embryogenesis in flax has been reviewed (Pretova and Obert, 2003) but knowledge regarding factors that affect and control ELS formation in this plant species is still insufficient and often contradictory.

Protoplasts can be isolated from most tissues of flax or linseed. Protoplasts have been widely used in a number of biochemical and physiological studies.

### **Genome mapping**

In the past few years, several types of molecular markers including random amplified polymorphics DNA (RAPDs), restriction fragment length polymorphisms (RFLPs), amplified fragment length polymorphisms (AFLPs), simple sequence repeats (SSRs) and EST-SSRs have been employed to analyze flax genetic diversity (Adugna et al., 2006; Cloutier, 2009). Sixty three QTLs for eighteen important agronomic traits have been recently identified (Vromans, 2006), providing new targets for manipulation using biotechnology.

### **Transformation techniques**

The first reports of the successful gene transfer by *Agrobacterium* into flax were relatively quickly after the original breakthrough in tobacco as early as 1986. The regeneration of transformed flax shoots via a callus stage has been described (Basiran et al., 1987). Regeneration of flax plants transformed by *Agrobacterium rhizogenes* was reported in 1988. However, the first successful verified transformation of flax using *Agrobacterium tumefaciens* and the consequent production of glyphosate-tolerant plants was described by Jordan and McHughen (1988) and successful uptake of a resistance gene to the herbicide sulphonylurea was reported by McSheffrey et al. (1992) and later an improved transformation procedure was published by Dong and McHughen (1993). However, Mlynarova et al. (1994) published a high efficiency method, used for a number of other studies. Rakousky et al. (1999) developed a transformation system based on the antibiotic selection agent hygromycin.

Though all methods to this date had been based on *Agrobacterium*-mediated transformation, Ling and Binding (1997) reported regeneration of transgenic plants through direct transformation of protoplasts with PEG and *A. tumefaciens* mediated transformation of plastocytes. Wijayanto and McHughen (1999) reported transformation of flax using particle bombardment and such an approach was adopted for transient expression studies of various seed-specific promoters. More applied transformation targets were reported by Ayliffe et al. (2002) investigating up-regulated genes at rust infection sites.

## **Oilseed crops**

## ***Brassica carinata* A. Braun**

### **Introduction**

Ethiopian or Abyssinian mustard (*Brassica carinata* A. Braun) is an amphidiploid with one genome from *Brassica nigra* L. Koch and the other from *Brassica oleracea* L. (genome BBCC,  $2n=34$ ).

With the additional agronomic advantage of its better tolerance to semi-arid conditions (Malik, 1990), the species has recently gained the interest of researchers in Canada and Spain (Raney et al., 1995; Velasco et al., 1995a). As appealing as it may be, from an agronomic standpoint, the stigma of low-quality oil from the seed of *B. carinata* is a legacy of its long-standing mediocrity. The reason for this is its high erucic acid content, which is reported to be in the range of  $35\pm 44\%$  (Becker et al., 1999). This is above the level acceptable, from a nutritional stand- point, in canola-quality rapeseed, which has now replaced all the traditional high-erucic acid types in Europe and Canada (Downey, 1990).

*Brassica carinata* is an adequate oil-bearing crop that is well-adapted to marginal regions (i.e., Andalusia (Spain), which is one of the poorest regions of the EU). Non food cultures in set-aside lands can significantly decrease the enormous amount of subsidies spent for agricultural overproduction in Europe, which leads to an increase in farmer incomes as well as the creation of new employment (Dorado et al., 2004). This crop is drought-resistant and grown in arid regions such as Andalusia.



**Figure 1:** A mature *Brassica carinata* plant

### **Biotechnology approaches**

#### **Inheritance of traits**

Ethiopian mustard possesses a number of agronomic advantages over other oilseed crops. The restricted amount of genetic variability available in natural *B. carinata* for traits like inbuilt resistance to drought, diseases and pests has constrained the breeding programmes aimed at improvement of the crop. Erucic acid is a trait which in high content is undesirable for use of *B. carinata* as a vegetable oil. Although efforts have been made to improve its quality, much has to be done to use natural variations that might exist within the species for fatty acid contents (Alemayehu and Becker, 2001).

A way to develop low erucic acid genotypes is through induced mutagenesis using ethyl methane sulphonate as the mutagen. It's been successful the developing diverse *B. carinata* mutant progenies with reduced erucic acid content, high oleic acid and high oil content which is a significant advance towards the development of Ethiopian mustard lines with canola characteristics (Sheikh et al., 2009).

The change of the properties of *Brassica carinata* can be achieved also through an efficient system of mutagenesis using ultraviolet (UV) light irradiation of isolated microspores (Barro et al., 2002).

### **Breeding achievements**

Microspore culture in combination with induced mutations can speed up breeding programmes, since homozygous doubled haploid (DH) lines exhibiting modified agronomic traits can be rapidly obtained (Maluszynski et al., 1995). Although microspore embryogenesis has been reported for *B. carinata*, information on differences in embryogenic response between genotypes is scanty. Regeneration has also been reported for cotyledons (Narasimhulu and Chopra, 1988) and hypocotyls. The frequency of plant regeneration from cultured cells and tissues of *B. carinata* has been relatively low in the past. But shoots of *B. carinata* can be regenerated at high frequency (100%) via selection of explants and manipulation of culture medium.

Among the economically important Brassica crops, plants have been produced from protoplasts of *B. oleracea* (Kao et al., 1990), *B. nigra* (Gupta et al., 1991), *B. juncea* (Kirti and Chopra, 1990), and *B. napus* (Thomzik and Haln 1990). In *B. carinata*, plants have been produced from hypocotyl protoplasts at a moderate frequency (Choung et al., 1987) and from cotyledon protoplasts of *B. carinata* using agarose embedding techniques (Jaiswal et al., 1990). Another approach involves high frequency regeneration from hypocotyl protoplasts of *B. carinata* without agarose embedding for three divergent genotypes (Narasimhulu et al., 1992).

### **Plant transformation**

Plant transformation systems have been developed for all the major Brassica species and also for *B. carinata* opening the way for genetic engineering to obtain transgenic plants with modified agronomic traits. Mutation techniques have been widely used to improve yield, disease and pest resistance in crops.

Although transgenic *B. carinata* plants have been produced at a relatively low frequency (1.5%) through *Agrobacterium*-mediated gene transformation (Narasimhulu et al., 1992), an efficient and reliable genetic transformation methodology for this species has been developed, using cotyledonary petiole and hypocotyl explants from *in vitro* grown seedlings.

#### **Improvement of bio-components of interest**

Genetic engineering of plants offers many opportunities for the agrochemical, food processing, and pharmaceutical industries to develop new products and manufacturing processes. There have been engineered *B. carinata* plants to express a gene for the protein hirudin, which is a potent thrombin inhibitor. *B. carinata* is chosen for these studies because of its low frequency of outcrossing, amenability to tissue culture techniques high frequency of multiple shoot regeneration per explant, and its transformation frequency using *Agrobacterium*-mediated gene transfer. When using *B. carinata* for genetic transformation, the co-suppression approach is attractive, because the high transformation rate in this species affords the opportunity to generate the high numbers of transgenic lines necessary to observe a co-suppression event.

#### **Molecular markers**

The genus *Brassica* contains a number of species of outstanding agronomical importance and, as a consequence, has received considerable attention in the context of crop improvement by both conventional plant breeding and biotechnological programmes. These approaches often require reliable chromosome identification and karyotypical analysis in order, for example, to assay chromosome variation following *in vitro* culture and regeneration, to characterise substitution or addition lines, or simply to integrate genetic and physical maps.

The number of 18S-5.8S-25S rDNA loci in *Brassica* diploids (*Brassica nigra*, *Brassica oleracea* and *Brassica campestris*) and allotetraploids (*Brassica carinata*, *Brassica juncea* and *Brassica napus*) was first described by the Maluszynska and Heslop-Harrison in 1993. Further investigations described more precisely the genomic distribution of rDNA sites on prometaphase and metaphase chromosomes, and later determined their transcriptional activity.

## ***Brassica napus* L.**

### **Introduction**

*Brassica napus*, also known as rape, oilseed rape, rapa, rapeseed and (in the case of one particular group of cultivars) canola, is a bright yellow flowering member of the family Brassicaceae (mustard or cabbage family). The name derives from the Latin for turnip, *rāpum* or *rāpa*, and is first recorded in English at the end of the 14th century. Older writers usually distinguished the turnip and rape by the adjectives *round* and *long* (-rooted) respectively.



Kingdom:	Plantae
Division:	Magnoliophyta
Class:	Magnoliopsida
Subclass:	<u>Dilleniidae</u>
Order:	Brassicales
Family:	Brassicaceae
Genus:	<i>Brassica</i>
Variety:	<b><i>B. napus</i></b>

**Figure 1.** *Brassica napus* (rapeseed)

Oilseed rape (rapeseed; *Brassica napus* L., genome AACC,  $2n = 38$ ) arises from spontaneous hybridization between turnip (*Brassica rapa*) (AA,  $2n = 20$ ) and cabbage (*Brassica oleracea*) (CC,  $2n = 18$ ). It is the most important oilseed crop in Europe and the second one over the world after soybean (*Glycine max*).

### **Biotechnology approaches**

#### **Regeneration and transformation techniques**

Organogenesis is an indispensable tool for plant regeneration using tissue culture techniques and for plant transformation. Regeneration of plants via organogenesis has been accomplished from various tissues such as cotyledons (Ono et al., 1994), hypocotyls (Yang et

al., 1991), peduncle segments (Eapen and George, 1997), leaves, thin cell layers of epidermal and subepidermal cells (Klimaszewska and Keller, 1985), roots (Xu et al., 1982), and protoplasts (Hu et al., 1999). However, hypocotyl segments remain the most desirable explants for tissue culture and have been used for most Brassica species because of their ability to regenerate.

Somatic embryogenesis, which has been the subject of increasing research in the genus, has become one of the most desired pathways in the regeneration of plants via tissue culture because it bypasses the necessity of time-consuming and costly manipulation of individual explants, which is a problem with organogenesis. Microspores or anthers have been somatic embryogenesis explants of choice in most Brassica species. Somatic embryos have been obtained from hypocotyls, protoplast-derived colonies, and immature cotyledons (Turgut et al., 1998) in *B. napus*.

Protoplast fusion allows the creation of hybrid and cybrid combinations of species that are sexually incompatible, thus facilitating the transfer of genes from a related, but sexually incompatible species, to another without genetic transformation. This technology has allowed not only intrageneric hybridizations, but the production of intergeneric hybrids and cybrids as well. Various desirable traits from the parents have been transferred to the hybrids and cybrids using this technology. One success of protoplast fusion has been the production of disease-resistant hybrids. Somatic hybrids that are resistant to bacterial soft rot have been produced by the fusion of *B. rapa* and *B. oleracea* protoplasts (Ren et al., 2000). Interspecific hybrids have been produced by fusing mesophyll protoplasts of *B. juncea* and *B. spinescens* (Kirti et al., 1991b). Protoplast fusion between *B. oleracea* and *Moricandia nitens*, a C3–C4 photosynthesis intermediate wild species, resulted in the production of intergeneric hybrids that expressed a gas-exchange character that was intermediate between the two parents.

### **Molecular markers**

Several markers have been used to assess the genetic fidelity of *in vitro*-grown plants such as isozymes, RFLPs and RAPDs. However, these markers are suboptimal for genetic identification, giving way to improved PCR fingerprinting technology. In *B. oleracea* var. botrytis, Leroy et al. (2000) have used inter-simple sequence repeat (ISSR) markers to analyze the genetic stability of somatic embryos derived from hypocotyls. They did not find any polymorphism between different regenerants. However, in cauliflower callus, out of the 224 calluses analyzed, six exhibited original patterns, and in one of these PCR patterns differed at four polymorphic loci. The most frequent primer used for detection of polymorphisms was (CAA) (Leroy et al., 2001).

A better knowledge of genetic determinism of oil content will be relevant for the breeders to control the genetic advance of the crop. By using different segregating rapeseed populations, recent studies reported the identification of numerous QTL (7 to 14 regions per



study) involved in the control of oil content, which is consistent with the polygenic determinism of the trait (Renard et al., 2006). Each of the QTL accounted for less than 10% of the total oil content variance (Renard et al, 2006, Bancroft et al, 2006). Some of these QTL coincided with loci controlling erucic acid content, suggesting that it is a major determinant for oil content in oilseed rape. Additive effects were shown to be the main factors controlling oil content (Renard et al, 2006, Bancroft et al, 2006), with individual additive effect of the different alleles ranging from 0.2 to 1.2%. In addition, strong environmental effects underlie variations in oil content (Turner et al, 2003). Yield-related traits (such as biomass, harvest index, plant architecture, adaptation, resistance to biotic and abiotic constraints) may also indirectly affect yield by affecting the yield-component traits or by other, unknown mechanisms. Increasing evidence suggests that “fine-mapped” quantitative trait loci (QTL) or genes identified as affecting crop yield involve diverse pathways, such as seed number (Burstin et al. 2007; Xie et al. 2008; Xing et al. 2008; Xue et al. 2008), seed weight (Shomura et al. 2008; Wang et al. 2008; Xing et al. 2008; Xue et al. 2008), flowering time (Xie et al. 2008; Xue et al. 2008), plant height (Xie et al. 2008; Xue et al. 2008), branching (Xing et al. 2008), biomass yield (Burstin et al. 2007), resistance and tolerance to biotic and abiotic stresses Warrington et al. 2008), and root architecture (Hochholdinger et al. 2008).

### Genetic transformation

Transformation systems have been developed in almost all the economically important species of Brassica such as *B. juncea* (Barfield and Pua, 1991), *B. napus* (Moloney et al., 1989), *B. rapa* (Radke et al., 1992), *B. oleracea* (De Block et al., 1989), *B. nigra* (Gupta et al., 1993), and *B. carinata*. Oil quality improvement has been an important target for Brassica transformation (Liu et al., 2001). Brassica oil is in great global demand and technology is available to custom-tune fatty acid profiles in seeds. Other target traits that were investigated for improvement through genetic transformation were insect resistance (Halfhill et al., 2001), salt tolerance (Pan et al, 2009) and male sterility (Jagannath et al., 2002).

#### LIST OF GENES OF IMPORTANCE RECENTLY INTRODUCED IN BRASSICA CROPS AND THEIR FUNCTIONS

Species	Gene introduced	Function	Reference
<i>Brassica napus</i> L.	crsI-1	Sulfonylurea resistance	Blackshaw et al., 1994
<i>Brassica napus</i> L.	Bxn	Bromoxynil resistance	Zhong et al., 1997
<i>Brassica napus</i> L.	d12-desaturase	Production of high g-linolenic acid	Liu et al., 2001
<i>Brassica napus</i> L.	Garm FatA1	Increase in enzyme activity towards acyl-acyl carrier protein (ACP)	Facciotti et al., 1999
<i>Brassica napus</i> L.	phbA, phbB, phbC, orbk1B, phbB, phbC	Production of poly( $\beta$ -hydroxybutyrate)(PHB)	Houmiel et al., 1999
<i>Brassica napus</i> L.	CrtB	Increase in carotenoid production	Shewmaker et al., 1999
<i>Brassica napus</i> L.	Truncated synthetic Bt Cry1A (c)	Resistance to diamondback moth and cabbage looper	Stewart et al., 1996; Halfhill et al., 2001
<i>B. rapa</i> ssp.	pekinensis Synthetic Bt Cry1c	Resistance to diamondback moth	Cho et al., 2001
<i>B. rapa</i> (syn. <i>B. campestris</i> ) ssp. <i>Parachinensis</i>	Synthetic Bt Cry1Ab, Cry1Ac	Resistance to diamondback moth	Xiang et al., 2000
<i>Brassica carinata</i>	OBHIRT (oleosin-hirudin) fusion protein	Production of hirudin	Chaudhary et al., 1998
Rutabaga ( <i>B. napo brasica</i> )	Bt Cry1A (c)	Resistance to cabbage caterpillar ( <i>Pieris rapae</i> )	Li et al., 1995
Broccoli ( <i>B. oleracea</i> L. var. <i>italica</i> )	Bt Cry1A (c)	Resistance to diamondback moth	Metz et al., 1995b; Cao et al., 1999
<i>Brassica oleracea</i> var. <i>capitata</i> (cabbage)	Bt Cry1Ab3	Resistance to diamondback moth larvae	Jin et al., 2000
<i>Brassica juncea</i>	Bacterial CodA	Enhanced salt and cold tolerance	Prasad et al., 2000
<i>Brassica juncea</i>	Barnase	Male sterility	Jagannath et al., 2001

## PERENNIAL GRASSES

### *Miscanthus* sp.

#### Introduction

The genus *Miscanthus* belongs to the tribe *Andropogoneae* in the family *Poaceae* and was first described by Andersson (1855). Is a perennial C4-grass and has its origins in the tropics and subtropics, but different species are found throughout a wide climatic range in East Asia. It is an environmentally benign plant that can be grown at a low level of fertiliser input, especially nitrogen (Lewandowski et al., 2000). *Miscanthus* biomass can be used as solid biofuel e.g. for co-combustion with coal (Wagenaar and Vandenheuvel, 1997), a source of industrial fibre for paper pulp (Cappelletto et al., 2000), insulation material, hard boards and plant potting mixtures and used whole for roof thatching (Kjeldsen et al., 1999).



**Figure 1.** Different *Miscanthus* sp. ecotypes and reproductive organs

Compared with other C-4 genera, miscanthus is more tolerant to the cool climate of north-west Europe (Beale & Long, 1995). Once established, miscanthus is harvested annually and in Denmark needs a rotation of minimum 10–12 yr in order to depreciate establishment costs (Parsby, 1996). The European investigations during the first decade were almost exclusively conducted with one genotype, the sterile, triploid hybrid *M. x giganteus* (Hodkinson & Renvoize, 2001). In northern Europe *M. x giganteus* was difficult to establish and had a rather poor combustion quality because it did not senesce, which delayed leaching of minerals from the crop during winter (Jørgensen, 1997; Venendaal *et al*, 1997). Therefore, the genetic base of miscanthus has been broadened in Europe by collecting and screening existing genotypes and by developing the breeding methods (Deuter & Abraham, 1998).

#### Biotechnology approaches

##### Genetic mapping

First molecular studies in *Miscanthus* used isozymes (Von Wuhlish et al. 1994) and AFLPs. The AFLP approach is widely used to analyze the genetic diversity of European species of *Miscanthus*. The technique is an adequate and powerful tool to evaluate genetic

diversification, to analyse the success of hybridizations and to find wrong classifications (Greef et al, 1997).

An “offspring cross” mapping strategy in combination with the random amplified polymorphic DNA (RAPD) assay has been used by Atienza et al (2002) in order to construct the first genetic map of the species *Miscanthus sinensis* ( $2n = 2x = 38$ ). In addition, 17–18S rRNA has been found useful for elucidating the ancient evolutionary history of angiosperm families or flowering plants (Chaw et al. 1997) owing to its conserved nature; and the internal transcribed spacer region (ITS) has been widely used for phylogenetic reconstruction at specific or generic levels (Baldwin 1992). Another way that was studied in order to collect and detect genetic variation in *Miscanthus* species is through single pollen grain polymerase chain reaction (PCR).

*M. sinensis* is the donor of two of the three genomes of *M x giganteus*. The Mendelian segregation of five microsatellite loci tested by Hernández et al (2001) illustrates the usefulness of these markers for *Miscanthus* mapping, and the high levels of variability and reproducibility associated with microsatellite markers will allow them to be used as anchor markers between genetic maps of *Miscanthus* and maize.

### **Breeding achievements**

Although vegetative propagation systems like rhizome division or *in vitro* axillary shoot propagation are available, the development of an efficient embryogenic cell culture system in *Miscanthus* is desirable. Embryogenic culture systems have already been established in the triploid hybrid *Miscanthus x ogiformis* Honda Giganteus’ (Lewandowski and Kahnt, 1993; Holme and Petersen, 1996). With the purpose of improving the callus culture system of *Miscanthus x ogiformis* Honda Giganteus’, proline was included in MS and N6 callus induction and suspension culture media. Effects of proline were investigated on embryogenic callus formation, growth of suspension cultures and plant regeneration (Bæksted Holme et al, 1997). Petersen (1997) demonstrated that plant regeneration from *Miscanthus x ogiformis* Honda ‘Giganteus’ could be improved considerably by adding low concentrations of BA during callus induction, compared to previous investigations (Holme and Petersen, 1996).

## ***Panicum virgatum* L.**

### **Introduction**

Switchgrass (*Panicum virgatum* L.) is a perennial C4 grass propagated by seed that can be established at low cost and risk and requires very low inputs while giving high biomass yields given on marginal soils. Since the early 1990s the crop has been developed as a model herbaceous energy crop for ethanol and electricity production in the USA and in Canada and it is also being considered as a paper pulp production feedstock. In Europe switchgrass has been introduced as a potential energy crop only recently. Several studies on growth and yield showed

encouraging results, and it is now clear that switchgrass can be cultivated both in North and South Europe. Additionally, economic analysis under different scenarios found switchgrass to be more profitable than other conventional crops e.g. maize or alfalfa. The crop has the potential to play a role in supporting policies to increase the use of durable products, reduce CO<sub>2</sub> emissions, utilise marginal and set aside lands and provide new economic activities for rural communities.

It is a self-incompatible and largely cross-pollinated species with a genome constitution varying from diploid to decaploid. Natural populations are broadly classified into two main ecotypes, lowland and upland, based on morphology and natural habitat. Lowland types are usually tetraploid with genetic composition of ( $2n = 4x = 36$ ) with a DNA content of approximately 3 pg. Most of the upland types are either hexaploid ( $2n = 6x = 54$ ) or octaploid ( $2n = 8x = 72$ ) with octaploid DNA content of 5.9 to 6.2 pg 2C-1 (Lu et al., 1998). In a species such as switchgrass there exists a great deal of phenotypic variation derived from latitudinal adaptation across its natural range and local adaptation to soil, temperature, and moisture conditions (Casler et al., 2007). It is still largely undomesticated thus large gains might be realized through fixation of beneficial alleles in breeding populations.

## **Biotechnology approaches**

### **Hybridization**

It has been shown that heterosis exists in switchgrass for agronomic traits including biomass yield (Vogel et al. 2007). Switchgrass is a cross-pollinated species, and cross pollination is enforced by a gametophytic self-compatibility system that is similar to the S-Z incompatibility system found in other Poaceae (Martinez-Reyna and Vogel, 2002). No allelopathy has been reported in switchgrass (Carroll and Somerville, 2009). The existence of gametophytic self-incompatibility in switchgrass may make it possible to develop switchgrass hybrid cultivars using the method illustrated in Figure 1. Improvements in tissue culture for clonally propagating plants, such as the node propagation method should make hybrids based on self-incompatibility commercially feasible (Martinez-Reyna and Vogel, 2008).

### **Genetic transformation**

Transformation of switchgrass cells has been obtained by PEG-mediated DNA uptake of protoplasts (Mazarei et al., 2008) and particle bombardment of immature inflorescence-derived embryogenic callus (Richards et al., 2001). Herbicide resistance and visual markers have been used to identify transformants. Switchgrass was reported being genetically transformed via *Agrobacterium* only the past few years. Recently, another transient transformation system the Fast Agro-mediated Seedling Transformation (FAST) has been developed providing a rapid, efficient and economical assay of gene function in intact plants with minimal manual handling and without dedicated device (Li et al., 2009). Actually, transformation projects with the aim to down-regulate genes in the lignin pathway are currently

underway in several grasses including switchgrass (Noble Foundation Press Release; URL: [http://www.noble.org/Press\\_Release/ForageImprovement/BiomassGrant/index.html](http://www.noble.org/Press_Release/ForageImprovement/BiomassGrant/index.html); USDA., 2006). Recently, it has been reported the successful engineering of switchgrass for the synthesis of polyhydroxybutyrate (PHB) which is high molecular weight polyester (Somleva et al., 2008), demonstrating that this high-yielding biomass crop is amenable to the complex metabolic engineering strategies necessary to produce high-value biomaterials with lignocellulose-derived biofuels.

### **Genome mapping**

ESTs and genomic microsatellites are being developed for switchgrass (Tobias et al., 2008) and should be a good source of molecular markers. 61,585 high-quality ESTs have been generated and seventy-three percent of the assembled consensus sequences could be aligned with the sorghum, indicating a high degree of similarity (Tobias et al., 2008). Chloroplast polymorphisms and random amplified polymorphic DNA (RAPD) markers have been used to evaluate diversity among cultivars and natural populations of switchgrass (Hultquist et al., 1996; Missaoui et al. 2006). Microsatellite markers developed from conserved grass (CG), tall fescue (TF) and switchgrass ESTs were assessed on parents and a subset of this mapping population (Saha et al. 2007). The genetic variability within and among 31 switchgrass populations obtained from Germplasm Resources Information Network (GRIN) has been assessed by EST-SSR markers.

### **Tissue culture technology**

Regeneration from cells or tissues cultured *in vitro* is a fundamental requirement for most applications of plant biotechnology. Tissue culture protocols for both direct and callus-mediated shoot regeneration and somatic embryogenesis are available for switchgrass. Switchgrass plants can be regenerated from mature caryopses, young leaf segments and *in vitro*-developed inflorescences through somatic embryogenesis and organogenesis (Alexandrova et al., 1996). However, the most effective approach seems to be the nodal culture that is possible to produce approximately 500 plantlets from one parent plant in 12 wk. Advanced regeneration techniques have been recently developed for switchgrass at the University of Tennessee, including production of flowers from tissue (node) culture (McLaughlin and Kszos, 2005).

## References

### *Beta Vulgaris* L

- Catusse J, Strub J-M, Job C, van Dorsselaer A, Job D (2008) Proteomewide characterization of sugarbeet seed vigor and its tissue specific expression. *Proc Natl Acad Sci USA* 105: 10262-10267.
- Dechyeva D, Gindullis F, Schmidt T (2003) Divergence of satellite DNA and interspersions of dispersed repeats in the genome of the wild beet *Beta procumbens*. *Chromosome Res* 11: 3–21.
- Graham MW, Craig S and Waterhouse PM (1997) Expression patterns of ascular-specific promoters RolC and Sh in transgenic potatoes and their use in engineering *PLRV*-resistant plants. *Plant Mol Biol* 33: 729-735.
- Hermann K, Meinhard J, Dobrev P, Linkies A, Pesek B, Hess B, Machácková I, Fischer U and Leubner-Metzger G (2007) 1-Aminocyclopropane-1-carboxylic acid and abscisic acid during the germination of sugar beet (*Beta vulgaris* L.): a comparative study of fruits and seeds. *J Exp Bot* 58: 3047-3060.
- Jacobs G, Dechyeva D, Wenke T, Weber B, and Schmidt T (2009) A BAC library of *Beta vulgaris* L. for the targeted isolation of centromeric DNA and molecular cytogenetics of *Beta* species. *Genetica* 135: 157–167.
- Jaeger GD, Scheffer S, Jacobs A, Zambre M, Zobell O, Goossens A, Depicker A and Angenon G (2002) Boosting heterologous protein production in transgenic dicotyledonous seeds using *Phaseolus vulgaris* regulatory sequences. *Nat Biotechnol* 20: 1265-1268.
- Kubis S, Heslop-Harrison JS and Schmidt T (1997) A family of differentially amplified repetitive DNA sequences in the genus *Beta* reveals genetic variation in *Beta vulgaris* subspecies and cultivars. *J Mol Evol* 44: 310–320.
- Lindsey K, Gallois P and Eady C (1991) Regeneration and transformation of sugar beet by *Agrobacterium tumefaciens*. In *Plant Tissue Culture Manual B7* Kluwer Academic Publishers; 1991:1-13.
- McGrath JM, Elawady A, El-Khishin D, Naegele RP, Carr KM and de los Reyes BG (2008) Sugar beet germination: Phenotypic selection and molecular profiling to identify genes involved in abiotic stress response. *Acta Horticulturae* 782: 35- 49.
- Menzel G, Dechyeva D, Wenke T, Holtgrawe D, Weisshaar B and Schmidt T. Diversity of a Complex Centromeric Satellite and Molecular Characterization of Dispersed Sequence Families in Sugar Beet (*Beta vulgaris*). *Ann Bot-London* 102: 521–530.
- Outchkourov NS, Peters J, De Jong J, Rademakers W and Jongsma MA (2003) The promoter-terminator of chrysanthemum rbcS1 directs very high expression levels in plants. *Planta* 216: 1003-1012.
- de los Reyes BG and McGrath JM (2003) Cultivar-specific seedling vigor and expression of a putative oxalate oxidase germin-like protein in sugar beet (*Beta vulgaris* L.). *Theor Appl Genet* 107: 54-61.
- Satoh M, Kubo T, Nishizawa S, Estiati A, Itchoda N et al. (2004) The cytoplasmic male-sterile type and normal type mitochondrial genomes of sugar beet share the same complement of genes of known function but differ in the content of expressed ORFs. *Mol. Genet. Genomics* 272: 247–256
- Stahl DJ, Kloos DU and Hehl R (2004) A sugar beet chlorophyll a/b binding protein promoter void of G-box like elements confers strong and leaf specific reporter gene expression in transgenic sugar beet. *BMC* 4: 31.
- Yamamoto MP, Kubo T and Mikami T (2005) The 59-leader sequence of sugar beet mitochondrial atp6 encodes a novel polypeptide that is characteristic of Owen cytoplasmic male sterility. *Mol Genet Genom* 273: 342–349.
- Zhang J, Van Toai T, Huynh L and Preiszner J (2000) Development of flooding-tolerant *Arabidopsis thaliana* by autoregulated cytokinin production. *Mol Breed* 6: 135-144.

## *Sorghum bicolor* L.

- Bowers J E, Abbey C, Anderson S et al. (2003) A high-density genetic recombination map of sequence-tagged sites for *Sorghum*, as a framework for comparative structural and evolutionary genomics of tropical grains and grasses. *Genetics* 165: 367–386.
- Buchanan CD, Lim S, Salzman RA et al. (2005) *Sorghum bicolor*'s transcriptome response to dehydration, high salinity and ABA. *Plant Mol Biol* 58: 699–720.
- Carrari F, Benech-Arnold R, Osuna-Fernandez R et al. (2003) Genetic mapping of the *Sorghum bicolor* *vp1* gene and its relationship with preharvest sprouting resistance *Genome* 46: 253–258.
- Casa M, Mitchell SE, Hamblin MT et al. (2005) Diversity and selection in sorghum: simultaneous analyses using simple sequence repeats, *Theor Appl Genet* 111:23–30.
- Duodu G, Taylor JRN, Belton PS and Hamaker BR (2003) Factors affecting sorghum protein digestibility. *J Cereal Sci* 38: 117–131.
- Feltus F A, Hart GE, Schertz K F et al. (2006) Genetic map alignment and QTL correspondence between inter- and intraspecific sorghum populations. *Theor and Appl Genet* 112: 1295–1305.
- Gaut BS (2002) Evolutionary dynamics of grass genomes. *New Phytol* 154:15–28.
- Hamblin T, Salas Fernandez MG, Casa AM, Mitchell SE, Paterson AH and Kresovich S (2005) Equilibrium processes cannot explain high levels of short and medium-range linkage disequilibrium in the domesticated grass *Sorghum bicolor*. *Genetics* 171: 1247–1256.
- Hass-Jacobus BL, Futrell-Griggs M, Abernathy B, Westerman R, Goicoechea JL, Stein J, Klein P, Hurwitz B, Zhou B, Rakhshan F, Sanyal A, Gill N, Lin J-Y, Walling JG, Zhong Luo M, Siva J, Ammiraju S, Kudrna D, Kim HR, Ware D, Wing RA, San Miguel P and Jackson SA (2006) Integration of hybridization-based markers (overgos) into physical maps for comparative and evolutionary explorations in the genus *Oryza* and in *Sorghum*. *BMC Genom* 7:199.
- Jessup et al, 2003
- Kim J-S, Klein PE, Klein RR, Price HJ, Mullet JE and Stelly DM (2005) Chromosome identification and nomenclature of *Sorghum bicolor*. *Genetics* 169: 1169–1173.
- Klein PE, Klein RR, Cartinhour SW et al. (2000) A high throughput AFLPbased method for constructing integrated genetic and physical maps: progress toward a sorghum genome map. *Genome Res* 10: 789–807.
- McIntyre CL, Tao Y, Jordan DR, Drenth J and Henzell RG (2001) In “Proceedings of the 4th Australia Sorghum Conference, Koorablyn, 5–8 February 2001” (A. K. Borrell and R. G. Henzell, Eds.), CD-ROM Format, ISBN 0-7242-2163-8.
- Menz MA, Klein RR, Mullet JE, Obert JA, Unruh NC and Klein PE (2002) A high density genetic map of *Sorghum bicolor* L. Moench based on 2926 AFLP, RFLP, and SSR markers. *Plant Mol. Biol.* 48: 483–499
- Paterson H, Lin Y-R, Li Z et al. (1995) Convergent domestication of cereal crops by independent mutations at corresponding genetic loci. *Science* 269: 1714–1718.
- Peterson DG, Schulze SR, Sciara EB et al. (2002) Integration of cot analysis, DNA cloning, and high-throughput sequencing facilitates genome characterization and gene discovery. *Genome Res* 12: 795–807.
- Pratt LH, Liang C, Shah M et al. (2005) Sorghum expressed sequence tags identify signature genes for drought, pathogenesis, and skotomorphogenesis from a milestone set of 16,801 unique transcripts. *Plant Physiol* 139: 869– 884.
- Salzman RA, Brady JA, Finlayson SA, Buchanan CD, Summer EJ, Sun F, Klein PE, Klein RR, Pratt LH, Cordonnier-Pratt M and Mullet JE (2005) Transcriptional Profiling of Sorghum Induced by Methyl Jasmonate, Salicylic Acid, and Aminocyclopropane Carboxylic Acid Reveals Cooperative Regulation and Novel Gene Responses. *Plant Physiol* 138:352–368.
- Ulanch PE, Childs KL, Morgan PW and Mullet JE (1996) Molecular markers linked to Ma(1) in sorghum. *Plant Physiol* 111: 709.
- Wen L, Tang HV, Chen W et al. (2002) Development and mapping of AFLP markers linked to the sorghum fertility restorer gene *rf4*. *Theor Appl Genet* 1044: 577–585.
- Winn JA, Mason RE, Robbins AL, Rooney WL and Hays DB (2009) QTL mapping of a High Protein Digestibility Trait in *Sorghum bicolor*. *International Journal of Plant Genomics*, Article ID 471853.

- Woo S-S, Jiang J, Gill B S, Paterson AH and Wing RA (1994) Construction and characterization of a bacterial artificial chromosome library of *Sorghum bicolor*. *Nucleic Acids Res* 22: 4922–4931.

## *Eucalyptus*

- Achere V, Faivre-Rampant P, Jeandroz S, Besnard G, Markussen T, Aragones A, Fladung M, Ritter E and Favre JM (2004) A full saturated linkage map of *Picea abies* including AFLP, SSR, ESTP, 5S rDNA and morphological markers. *Theor Appl Genet* 108:1602-1613.
- Andrew RL, Peakall R, Wallis IR, Wood JT, Knight EJ and Foley WJ (2005) Marker-Based Quantitative Genetics in the Wild?: The Heritability and Genetic Correlation of Chemical Defenses in *Eucalyptus*. *Genetics* 171: 1989–1998.
- Bandyopadhyay S, Cane K, Rasmussen G and Hamill JD (1999) Efficient plant regeneration from seedling explants of two commercially important temperate eucalypt species *Eucalyptus nitens* and *E. globulus*. *Plant Sci* 140:189–198.
- Bandyopadhyay S and Hamill JD (2000) Ultrastructural studies of somatic embryos of *Eucalyptus nitens* and comparisons with zygotic embryos found in mature seeds. *Ann Bot* 86: 237–244.
- Bossinger G and Leitch M (2000) Isolation of cambium-specific genes from *Eucalyptus globulus* Labill. In: *molecular biology of wood formation*. BIOS Scientific, Oxford, pp 203–207.
- Brondani RPV, Brondani C, Tarchini R and Grattapaglia D (1998) Development and mapping of microsatellite based markers in *Eucalyptus*. *Theor App Genet* 97: 816-829.
- Brondani RPV, Williams ER, Brondani C and Grattapaglia D (2006) A microsatellite-based consensus linkage map for species of *Eucalyptus* and a novel set of 230 microsatellite markers for the genus. *BMC Plant Biology* : 620.
- Brondani RPV and Grattapaglia D (2002) Towards the construction of a genus wide reference linkage map for *Eucalyptus* based on microsatellite markers. *Mol. Gen. Genomics* 267: 338-347.
- Brondani RPV, Williams ER, Brondani C and Grattapaglia D (2006) A microsatellite-based consensus linkage map for species of *Eucalyptus* and a novel set of 230 microsatellite markers for the genus. *BMC Plant Biology* : 620.
- Brown G, Gill G, Sewell M, Wheeler N, Megraw R and Neale D (2001) Towards association studies in forest trees: wood property QTL verification, candidate genes, and SNPs in Loblolly pine (*Pinus taeda*). *Abstracts of the 9th Joint Conifer Biotechnology Working Group and IUFRO Meeting: “Wood, Breeding, Biotechnology and Industrial Expectations”*, Bordeaux, France, p:104.
- Bundock PC, Hayden M and Vaillancourt RE (2000) Linkage maps of *Eucalyptus globulus* using RAPD and microsatellite markers. *Silvae Genet* 49: 223–232.
- Carbonnier L (2004) The future of *Eucalyptus* plantations. In: Borralho N, Pereira J, Marques C, Coutinho J, Madeira M, TomuM(eds) *IUFRO on silviculture and improvement of eucalypts: Eucalyptus in a changing world*. Raiz Instituto, Aveiro, p 29.
- Costa da Cruz M, Gomes Caldas D, Tozelli Carneiro R, Moon DH, Salvatierra G, Franceschini LM, de Andrade A, Fiorani Celedon PA, Oda S and Labate CL (2008) SAGE transcript profiling of the juvenile cambial region of *Eucalyptus grandis*. *Tree Physiol* 28: 905-919.
- Dale GT, Aitken K and Sasse J (2000) Development of salt-tolerant *E. camaldulensis* 3 *E. grandis* hybrid clones using phenotypic selection and genetic mapping. In: Dungey H.S., Dieters M.J. and Nikles D.G. (eds), *Hybrid Breeding and Genetics of Forest Trees*. Proceedings of QFRI/CRC-SPF Symposium, 9– 14th April 2000 Noosa, Queensland, Australia. Department of Primary Industries, Brisbane, pp. 227–233.
- Dhawan V and Saxena S (2004) *Cloning Forestry Species*. Plant Biotechnology and Molecular Markers. Anamaya Publishers, New Delhi, India.
- Fullard K and Moran G (2003) Identification of frost tolerance genes in a *Eucalyptus* hybrid cross. In: Sundberg B (ed) *IUFRO tree biotechnology*. Umea Plant Science Centre, Umea, pp S6–S18.
- Grattapaglia D (2000) Molecular breeding of *Eucalyptus*: state of the art, applications and technical challenges. In: *Molecular Markers and Genome Mapping in Woody Plants* (Jain, S.M. and Minocha, S.C., eds.). Vol. 1. Kluwer Academic Publishers, Dordrecht, The Netherlands, pp. 451-474.
- Grattapaglia D (2004) Integrating genomics into *Eucalyptus* breeding. *Genet Mol Res* 3: 369-379



- Junghans D, Alfenas AC, Brommonschenkel SH, Oda S, Mello EJ and Grattapaglia D (2003) Resistance to Rust in *Eucalyptus*: mode of inheritance and mapping of a major gene with RAPD markers. *Theor Appl Genet* 108: 175-180.
- Kirst M, Myburg AA, De Leon JPG, Kirst ME, Scott J and Sederoff R (2004) Coordinated Genetic Regulation of Growth and Lignin Revealed by Quantitative Trait Locus Analysis of cDNA Microarray Data in an Interspecific Backcross of *Eucalyptus*. *Plant Physiol* 135: 2368–2378.
- Komulainen P, Brown GR, Mikkonen M, Karhu A, Garcia-Gil MR, O'Malley D, Lee B, Neale DB, Savolainen O (2003) Comparing ESTbased genetic maps between *Pinus sylvestris* and *Pinus taeda*. *Theor Appl Genet* 107: 667-678.
- Krutovsky KV, Troggio M, Brown GR, Jermstad KD, Neale DB (2004) Comparative mapping in the Pinaceae. *Genetics* 168: 447-461.
- Külheim C, Yeoh SH, Maintz J, Foley WJ and Moran GF (2009) Comparative SNP diversity among four *Eucalyptus* species for genes from secondary metabolite biosynthetic pathways. *BMC Genomics* 10: 452.
- McKinnon GE, Vaillancourt RE, Tilyard PA and Potts BM (2001) Maternal inheritance of the chloroplast genome in *Eucalyptus globulus* and interspecific hybrids. *Genome* 44: 831–835.
- Myburg A, Griffin R, O'Malley D, Sederoff RR and Whetten R (2001) Genetic analysis of growth and wood quality traits in interspecific backcross families of *Eucalyptus grandis* and *Eucalyptus globulus*. *Plant and Animal Genome IX Conference*, San Diego, CA, USA, Abstract W82.
- Novaes E, Drost DR, Farmerie WG, Pappas GJ, Grattapaglia D, Sederoff RR and Kirst M (2008) High-throughput gene and SNP discovery in *Eucalyptus grandis*, an uncharacterized genome. *BMC Genomics* 9: 312.
- Rengel D, San Clemente H, Servant F, Ladouce N, Paux E, Wincker P, Couloux A, Sivadon P and Grima-Pettenati J (2009) A new genomic resource dedicated to wood formation in *Eucalyptus*. *BMC Plant Biology* 9: 36.
- Shepherd M and Jones ME (2005) Molecular markers in tree improvement: characterisation and use in *Eucalyptus*, pp. 399–409, in *Molecular marker systems in plant breeding and crop improvement*, edited by H. Lorz & G. Wenzel. Springer-Verlag, Heidelberg, Germany.
- Thamarus K, Groom K, Murrell J, Byrne M and Moran G (2002) A genetic linkage map for *Eucalyptus globulus* with candidate loci for wood, fibre and floral traits. *Theor Appl Genet* 104: 379-387.
- Thumma BR, Nolan MF, Evans R and Moran GF (2005) Polymorphisms in cinnamoyl coa reductase (ccr) are associated with variation in microfibril angle in *Eucalyptus* spp. *Genetics* 171: 1257-1265.
- Vaillancourt RE, Petty A and McKinnon GE (2004) Maternal inheritance of mitochondria in *Eucalyptus globulus*. *J Hered* 95: 353–355.
- Yin TM, DiFazio SP, Gunter LE, Riemenschneider D and Tuskan GA (2004) Large-scale heterospecific segregation distortion in *Populus* revealed by a dense genetic map. *Theor Appl Genet* 109: 451-463.
- Zhou Y, Gwaze DP, Reyes-Valdes MH, Bui T and Williams CG (2003) No clustering for linkage map based on low-copy and undermethylated microsatellites. *Genome* 46: 809-816.

### ***Populus spp***

- Arisi A-CM, Noctor G, Foyer CH, Jouanin L (1997) Modification of thiol contents in poplars (*Populus tremula*3 *P. alba*) overexpressing enzymes involved in glutathione synthesis. *Planta* 203: 362–372.
- Arnaud D, Dejardin A, Leple JC, Legase-Descauses MC and Pilate G (2007) Genome-Wide Analysis of LIM Gene Family in *Populus trichocarpa*, *Arabidopsis thaliana*, and *Oryza sativa*. *DNA Res* 14: 103–116.
- Bhalerao R, Keskitalo J, Sterky F, Erlandsson R, Bjorkbacka H, Jonsson Birve S, Karsson J, Gardstrom P, Gustafsson P, Lundeberg J and Jansson S (2003) Gene expression in autumn leaves. *Plant Physiol* 131: 430-442.
- Bogeat-Triboulot MB, Brosche M, Renaut J, Jouve L, Le Thiec D, Fayyaz P, Vinocur B, Witters E, Laukens K, Altman A, Hausman JF, Polle A, Kangasjarvi J and Dreyer E (2007) Gradual soil water depletion results in reversible changes of gene expression, protein profiles, ecophysiology, and growth performance in *Populus euphratica*, a poplar growing in arid regions. *Plant Physiol* 143: 876-892.

- Brosche M, Vinocur B, Alatalo ER, Lamminmaki A, Teichmann T et al. (2005) Gene expression and metabolite profiling of *Populus euphratica* growing in the Negev desert. *Genome Biol.* 6, R101.
- Daniell H (2007) Transgene containment by maternal inheritance: effective or elusive? *Proc Natl Acad Sci USA* 104: 6879–6880.
- Eriksson ME, Israelsson M, Olsson O and Moritz T (2000) Increased gibberellins biosynthesis in transgenic trees promotes growth, biomass production and xylem fiber length. *Nat. Biotechnol.* 18: 784–788.
- Ferreira S, Hjerno K, Larsen M, Wingsle G, Larsen P, Fey S, Roepstorff P and Pais MS (2006) Proteome profiling of *Populus euphratica* Oliv. Upon heat stress. *Ann Bot* 98: 361–377.
- Hertzberg M, Aspeborg H, Schrader J, Andersson A, Erlandsson R, Blomqvist K, Bhalariao R, Uhlen M, Teeri TT, Lundeberg J et al. (2001) A transcriptional roadmap to wood formation. *Proc Natl Acad Sci USA* 98: 14732–14737.
- Kalluri UC, DiFazio SP, Brunner AM and Tuskan GA (2007) Genome-wide analysis of *Aux/IAA* and *ARF* gene families in *Populus trichocarpa*. *BMC Plant Biology* 7: 59.
- Leple J-C, Dauwe R, Morreel K, Storme V, Lapierre C et al. (2007) Downregulation of cinnamoyl-coenzyme A reductase in poplar: multiple-level phenotyping reveals effects on cell wall polymer metabolism and structure. *Plant Cell* 19: 3669–3691.
- Lindroth RL and Hwang SY (1996) Diversity, redundancy, and multiplicity in chemical defense systems of aspen. In JT Romeo, JA Saunders, P Barbosa, eds, *Phytochemical Diversity and Redundancy in Ecological Interactions*. Plenum Press, New York, pp 25–56.
- Liu J, Hu D, Jiang G and Schnoor JL (2009) In Vivo Biotransformation of 3,3',4,4'-Tetrachlorobiphenyl by Whole Plants-Poplars and Switchgrass. *Environ. Sci. Technol.*, 43: 7503–7509.
- Loivamki M, Louis S, Cinege G, Zimmer I, Fischbach RJ and Schnitzler JP (2007) Circadian Rhythms of Isoprene Biosynthesis in Grey Poplar Leaves. *Plant Physiol* 143: 540–551.
- Major IT and Constabel CP (2006) Molecular analysis of poplar defense against herbivory: comparison of wound- and insect elicitor-induced gene expression. *New Phytol* 172:617–635.
- Mellway RD, Tran LT, Prouse MB, Campbell MM and Constabel CP (2009) The Wound-, Pathogen-, and Ultraviolet B-Responsive MYB134 Gene Encodes an R2R3 MYB Transcription Factor That Regulates Proanthocyanidin Synthesis in Poplar. *Plant Physiol* 150: 924–941.
- Miranda M, Ralph SG, Mellway R, White R, Heath MC, Bohlmann J and Constabel CP (2007) The transcriptional response of hybrid poplar (*Populus trichocarpa* X *P. deltoides*) to infection by *Melampsora medusae* leaf rust involves induction of flavonoid pathway genes leading to the accumulation of proanthocyanidins. *Mol Plant-Microbe Interac* 20: 816–831.
- Moreau C, Aksenov N, Lorenzo MG, Segerman B, Funk C, Nilsson P, Jansson S and Tuominen H (2004) A genomic approach to investigate developmental cell death in woody tissues of *Populus* trees. *Genome Biol* 6: R34.
- Nicole MC, Hamel LP, Morency MJ, Beaudoin N, Ellis BE and Seguin A (2006) MAP-ping genomic organization and organ-specific expression profiles of poplar MAP kinases and MAP kinase kinases. *BMC Genomics* 7: 223.
- Noctor G, Arisi A-CM, Jouanin L and Foyer CH (1998) Manipulation of GSH and amino acid biosynthesis in the chloroplast. *Plant Physiol* 118: 471–482.
- Ogata Y, Suzuki H and Shibata D (2009) A database for poplar gene coexpression analysis for systematic understanding of biological processes, including stress responses. *J Wood Sci* 55: 395–400.
- Okumura S, Sawada M, Park YW, Hayashi T, Shimamura M, Takase H and Tomizawa K (2006) Transformation of poplar (*Populus alba*) plastids and expression of foreign proteins in tree chloroplasts. *Transgenic Res* 15: 637–646.
- Pavy N, Johnson JJ, Crow JA, Paule C, Kunau T, MacKay J and Retzel EF (2006) ForestTreeDB: a database dedicated to the mining of tree transcriptomes. *Nuclei Acid Res* 35: D888–D894.
- Poke FS, Potts BM, Vaillancourt RE and Raymond CA (2006) Genetic parameters for lignin, extractives and decay in *Eucalyptus globulus*. *Ann For Sci* 63: 813–821.
- Rae AM, Pinel MPC, Bastien C, Sabatti M, Street NR, Tucker J, Dixon C, Marron N, Dillen SY and Taylor G (2008) QTL for yield in bioenergy *Populus*: identifying GxE interactions from growth at three contrasting sites. *Tree Genet Genomes* 4: 97–112.
- Rae AM, Street NR, Robinson KM, Harris N and Taylor G (2009). Five QTL hotspots for yield in short rotation coppice bioenergy poplar: The Poplar Biomass Loci. *BMC Plant Biol* 9: 23.

- Ralph SG, Chun HJE, Cooper D, Kirkpatrick R, Kolosova N, Gunter L, Tuskan GA, Douglas CJ, Holt RA, Jones SJM, Marra MA and Bohlmann J (2008) Analysis of 4,664 high-quality sequence-finished poplar full-length cDNA clones and their utility for the discovery of genes responding to insect feeding. *BMC Genomics* 9: 57.
- Ramirez-Carvajal GA, Morse AM and Davis JM (2008) Transcript profiles of the cytokinin response regulator gene family in *Populus* imply diverse roles in plant development. *New Phytol* 177: 77–89.
- Schrader J, Nilsson J, Mellerowicz E, Berglund A, Nilsson P, Hertzberg M and Sandberg G (2004) A High-Resolution Transcript Profile across the Wood-Forming Meristem of Poplar Identifies Potential Regulators of Cambial Stem Cell Identity. *Plant Cell* 16: 2278–2292.
- Stein L (2001) Genome annotation: from sequence to biology. *Nat Rev Genetics* 2: 493–503.
- Sterky F, Bhalariao RR, Unneberg P, Segerman B, Nilsson P, Brunner AM, Charbonnel-Campaa L, Jonsson-Lindvall J, Tandere K, Strauss SH, et al. (2004) A *Populus* EST resource for plant functional genomics. *Proc Natl Acad Sci USA* 101: 13951–13956.
- Tuskan GA, DiFazio S, Jansson S, Bohlmann J, Grigoriev I et al. (2006) The genome of black cottonwood, *Populus trichocarpa* (Torr. & Gray). *Science* 313: 1596–1604.
- Waters ER, Aevermann BD and Sanders-Reed Z (2008) Comparative analysis of the small heat shock proteins in three angiosperm genomes identifies new subfamilies and reveals diverse evolutionary patterns. *Cell Stress Chaperones* 13: 127–142.
- Whitham TG, Bailey JK, Schweitzer JA, Shuster SM, Bangert RK, Leroy CJ, Lonsdorf EV, Allan GJ, DiFazio SP, Potts BM, et al. (2006) A framework for community and ecosystem genetics: from genes to ecosystems. *Nat Rev Genet* 7: 510–523.
- Wilkins O, Nahal H, Foong J, Provart NJ and Campbell MM (2009) Expansion and diversification of the *Populus* R2R3-MYB family of transcription factors. *Plant Physiol* 149: 981–993.
- Wullschlegel SD, Yin TM, DiFazio SP, Tschaplinski TJ, Gunter LE, Davis MF and Tuskan GA (2005) Phenotypic variation in growth and biomass distribution for two advanced-generation pedigrees of hybrid poplar. *Can J For Res* 35: 1779–1789.
- Zsuffa L (1975) A summary review of interspecific breeding in the genus *Populus* L. In: *Proceedings, 14th Meeting of the Canadian Tree Improvement Association, Part 2*. Department of Environment, Canadian Forestry Service, Ottawa, Canada, pp. 107–123.

### *Cynara cardunculus* L.

- Acquadro A, Portis E, Lee D, Donini P and Lanteri S (2005a) Development and characterization of microsatellite markers in *Cynara cardunculus* L. *Genome* 48: 217–225.
- Acquadro A, Portis E, Albertini E et al. (2005b) M-AFLP-based protocol for microsatellite loci isolation in *Cynara cardunculus* L. (Asteraceae). *Molecular Ecology Notes* 5: 272–274.
- Acquadro A, Lanteri S, Scaglione D, Arens P, Vosman B, Portis E (2009) Genetic mapping and annotation of genomic microsatellites isolated from globe artichoke. *Theor Appl Genet* 118:1573–1587. Arce et al., 2004
- Basnizki J and Zohary D (1994) Breeding of seed planted artichoke. *Plant Breed Rev* 12:253–269.
- Comino C, Hehn A, Moglia A, Menin B, Bourgaud F, Lanteri S and Portis E (2009) The isolation and mapping of a novel hydroxycinnamoyltransferase in the globe artichoke chlorogenic acid pathway. *BMC Plant Biology* 2009, 9:30 [DOI:10.1186/1471-2229-9-30].
- De Paolis A, Pignone D, Morgese A and Sonnante G (2008) Characterization and differential expression analysis of artichoke phenylalanine ammonia-lyase-coding sequences. *Physiol Plant* 132: 33–43.
- Esteva, J and Martínez, J (2004) Evaluation of yield, earliness and head characteristics of bull variant plants in globe artichoke varieties ‘Blanca de Tudela’ and ‘Violet de Provence’ at Murcia. *Acta Hort* 660: 117–121.
- Guarino C, De Simone L, Santoro S, Caira S, Lilla S, Calabrese MG, Chianese L and Addeo F (2010) The Proteomic Changes in *Cynara Cardunculus* L. var. *altilis* DC Following the Etiolation Phenomena Using *De Novo* Sequence Analysis. *J Bot* [DOI:10.1155/2010/496893].
- Keurentjes J, Fu J, de Vos C, Lommen A, Hall R, Bino R, van der Plas L, Jansen R, Vreugdenhil D, Koornneef M (2006) The genetics of plant metabolism. *Nat Genet* 38: 842–849.

- Lanteri S, Saba E, Cadinu M, Mallica GM, Baghino L and Portis E (2004a) Amplified fragment length polymorphism for genetic diversity assessment in globe artichoke. *Theor Appl Genet* 108:1534–1544.
- Lanteri S, Acquadro A, Saba E and Portis E (2004b) Molecular fingerprinting and evaluation of genetic distances among selected clones of globe artichoke (*Cynara scolymus* var. *cardunculus* L.) ‘Spinoso sardo’. *J. Hort. Sci. Biotech* 79: 863-870.
- Lanteri S and Portis E (2008) Globe Artichoke and Cardoon. *Handbook of Plant Breeding*, vol 1, part 1: 49-74 [DOI: 10.1007/978-0-387-30443-4\_]
- Mauromicale G and Ierna A (2000) Characteristics of Head of Seed-Grown Globe Artichoke (*Cynara cardunculus* L. var. *scolymus* (L.) Fiori) as effected by Harvest Period, Sowing Date and Gibberellic Acid. 20: 197-204.
- Miguel A, Baixauli C, Aguilar JM, Giner A, Maroto JV, Lopez S, San Bautista A and Pascual B (2004) Gibberellic Acid Concentrations in Seed Propagated Artichoke. V International Congress on Artichoke 5-8 May, 2003. Tudela- Navarra, Spain. 167-172.
- Moglia A, Comino C, Portis E, Acquadro A, De Vos RCH, Beekwilder J and Lanteri S (2009) Isolation and mapping of a C3'H gene (CYP98A49) from globe artichoke, and its expression upon UV-C stress. *Plant Cell Rep* 28: 963–974.
- Motzo R and Deidda M (1993) Anther and ovule culture in globe artichoke. *J Genet Breed* 47: 263-266.
- Pagnotta MA and Noorani A (2008) European genetic resources of *Cynara* spp. - the CYNARES Project. *Bioversity Newsletter for Europe* issue No. 36: 11.
- Pecaut P (1983) Amelioration des varietes d'artichaut: varietes e multiplication vegetative, varietes e multiplication par semences, clones sans virus issus de multiplication *in vitro*. *Proces-verbal de la Seance de 12 Janvier, Acad Agric Fr*, pp 69–78.
- Portis E, Acquadro A, Comino C, Mauromicale G, Saba E and Lanteri S (2005a) Genetic structure of island populations of wild cardoon [*Cynara cardunculus* L. var. *sylvestris* (Lamk) Fiori] detected by AFLPs and SSRs. *Plant Science* 169: 199-210.
- Portis E, Barchi L, Acquadro A, Macua JI and Lanteri S (2005b) Genetic diversity assessment in cultivated cardoon by AFLP (amplified fragment length polymorphism) and microsatellite markers. *Plant Breeding* 124: 299-304.
- Raccuia SA, Mainolfi A, Mandolino G and Melilli MG (2004) Genetic diversity in *Cynara cardunculus* revealed by AFLP markers: comparison between cultivars and wild types from Sicily. *Plant Breeding* 123: 280–284.
- Rajjou L, Belghazi M, Huguet R, Robin C, Moreau A, Job C and Job D (2006) *Plant Physiol.* 141: 910–923.
- Rottenberg A and Zohary D (1996) The wild ancestry of the cultivated artichoke. *Genet Resour Crop Ev* 43: 53–58. Saccardo et al., 2007
- Scaglione D, Acquadro A, Portis E, Taylor CA, Lanteri S and Knapp SJ (2009) Ontology and diversity of transcript-associated microsatellites mined from a globe artichoke EST database. *BMC Genomics* 10:454 [doi:10.1186/1471-2164-10- 454].
- Sonnante G, De Paolis D, Lattanzio V and Perrino P (2002) Genetic variation in wild and cultivated artichoke revealed by RAPD markers. *Genet Resources Crop Evol* 49:247-252.
- Sonnante G, Cattonaro F, Scalabrin S , De Paola D, Pignone D and Morgante M (2011) First Glance Into *Cynara cardunculus* Genome By Next Generation Sequencing. Poster: Genome Sequencing & ESTs P041, Plant & Animal Genome XIX Conference, San Diego, California, USA.
- Stamigna C, Crinò P, Chatelet P and Saccardo F (2004). Induction of embryogenesis in isolated microspores of artichoke (*Cynara scolymus* L.). *Acta Hort* 660: 139-14.
- Thiellement H, Zivy M and Plomion C (2002) *J Chromatogr B* 782: 137–149.

### ***Cannabis sativa* L.**

- Alghanim HJ and Almirall JR (2003) Development of microsatellite markers in *Cannabis sativa* for DNA typing and genetic relatedness analyses. *Anal Bio Chem* 376: 1225–1233.
- Faeti V, Mandolino G and Ranalli P (1996) Genetic diversity of *Cannabis sativa* germplasm based on RAPD markers. *Plant Breed* 115: 367–370.
- Flachowsky H, Schumann E, Weber WE and Peil A (2001) Application of AFLP for the detection of sex-specific markers in hemp. *Plant Breed* 120: 305–309.

- Forapani S, Carboni A, Paoletti C, Moliterni VMC, Ranalli P et al. (2001) Comparison of hemp (*Cannabis sativa* L) varieties using RAPD markers. *Crop Sci* 41: 1682–1689.
- Hammond C T and Mahlberg PG (1977) Morphogenesis of capitate glandular hairs of *Cannabis sativa* (Cannabaceae). *Am J Bot* 64: 1023–1031.
- Hewavitharana AK, Golding G, Tempany G, King G and Holling N (2005) Quantitative GC–MS Analysis of  $\Delta^9$ -Tetrahydrocannabinol in Fiber Hemp Varieties. *J Anal Toxicol* 29: 258–261.
- Hillig KW and Mahlberg PG (2004) A chemotaxonomic analysis of cannabinoid variation in *Cannabis* (Cannabaceae). *Am J Bot* 91: 966–75.
- Kostic M, Pejic B and Skundric P (2008) Quality of chemically modified hemp fibers. *Bioresource Technol* 99: 94–99.
- Lu X and Clarke RC (1995) The cultivation and use of hemp (*Cannabis sativa* L.) in ancient China. *J International Hemp Association* 2: 26–30.
- Mandolino G and Carboni A (2004) Potential of marker-assisted selection in hemp genetic improvement. *Euphytica* 140: 107–120.
- de Meijer EPM, van der Kamp HJ and van Eeuwijk FA (1992) Characterisation of *Cannabis* accessions with regard to cannabinoid content in relation to other plant characters. *Euphytica* 62: 187–200.
- Rustichelli C, Ferioli V, Baraldi M, Zanoli P and Gamberini G (1998) Analysis of cannabinoids in fibre hemp plant varieties (*Cannabis sativa* L.) by high performance liquid chromatography. *Chromatographia* 47: 215–222.
- Small E and Beckstead HD (1973) Common cannabinoid phenotypes in 350 stocks of *Cannabis*. *Lloydia* 36: 144–165.
- de Zeeuw RA, Malingre M and Merkus FWHM (1972) Tetrahydrocannabinolic acid, an important component in the evaluation of *Cannabis* products. *J Pharm Pharmacol* 24: 1–6.

### ***Hibiscus cannabinus* L.**

- Banks SW, Gossett DR, Lucas MC, Milhollon EP and LaCell MG (1993) *Agrobacterium* mediated transformation of kenaf (*Hibiscus cannabinus* L.) with the  $\beta$ - glucuronidase (GUS) gene. *Plant MolBiol Rep* 11: 101–104.
- Kobaisy M, Tellez MR, Webber CL, Dayan FE, Schrader KK and Wedge DE (2001) Phytotoxic and Fungitoxic Activities of the Essential Oil of Kenaf (*Hibiscus cannabinus* L.) Leaves and Its Composition. *J Agric Food Chem* 49: 3768–3771.
- Kojima M, Shioiri H, Nogawa M, Nozue M, Matsumoto D, Wada A, Saiki Y and Kiguchi K (2004) *In planta* transformation of kenaf plants (*Hibiscus cannabinus* L.) by *Agrobacterium tumefaciens*. *J Biosc Bioeng* 98: 136–139.
- Liang XZ, Ding SW and Wong SM (2002) Development of a kenaf (*Hibiscus cannabinus* L.) protoplast system for a replication study of *Hibiscus* chlorotic ringspot virus. *Plant Cell Rep* 20: 982–986.
- Reichert NA. and Liu DL (1996) Protoplast isolation, culture, and fusion protocols for kenaf (*Hibiscus cannabinus*). *Plant Cell Tiss Org* 44: 201–210.
- Ruotolo G, Di Matteo A, Chialese P and Filippone E (2007) Cloning of kenaf (*Hibiscus cannabinus* l.) major lignin and cellulose biosynthesis gene sequences and their expression analysis during plant development. *Proceedings of the 51<sup>st</sup> Italian Society of Agricultural Genetics Annual Congress*. ISBN 978-88-900622- 7-8.
- Sanford JC, Smith FD and Russell JA (1993) Optimizing the biolistic process for different biological applications. *Methods Enzymol* 217: 483–509.
- Srivatanakul M, Park SH, Salas MG and Smith RH (2001) Transformation parameters enhancing T-DNA expression in kenaf (*Hibiscus cannabinus* L.) *J Plant Physiol* 158: 255–260.

### ***Linum usitatissimum* L.**

- Adugna W, Labuschagne MT and Viljoen CD (2006) The use of morphological and AFLP markers in diversity analysis of linseed. *Biodivers Conserv* 15:3193–3205.
- Ayliffe MA, Roberts JK, Mitchell HJ, Zhang R, Lawrence GJ, Ellis JG and Pryor TJ (2002) A plant gene up-regulated at rust infection sites. *Plant Physiol* 129: 169–180.
- Basiran N, Armitage P, Scott RJ and Draper J (1987) Genetic transformation of flax (*Linum usitatissimum*) by *Agrobacterium tumefaciens* regeneration of transformed shoots via a callus phase. *Plant Cell Rep* 6: 396–399.

- Bonell ML and Lassaga SL (2002) Genetic analysis of the response of linseed (*Linum usitatissimum* L.) somatic tissue to *in vitro* cultivation. *Euphytica* 125: 367–372.
- Bretagne B, Chupeau MC, Chupeau Y and Fouilloux G (1994) Improved flax regeneration from hypocotyls using thidiazuron as a cytokinin source. *Plant Cell Rep* 14: 120–124
- Bretagne-Sagnard B, Fouilloux G & Chupeau Y (1996) Induced albina mutations as a tool for genetic analysis and cell biology in flax (*Linum usitatissimum*). *J Exp Bot* 47: 189–194.
- Cloutier S, Niu Z, Datla R and Duguid S (2009) Development and analysis of ESTSSRs for flax (*Linum usitatissimum* L.). *Theor Appl Genet* 19: 53–63.
- Cunha ACG and Ferreira MF (1996) Somatic embryogenesis, organogenesis and callus growth kinetics of flax. *Plant Cell Tiss Org* 47: 1–8.
- Cunha A and Ferreira MF (1997) Differences in free sterols content and composition associated with somatic embryogenesis, shoot organogenesis and calli growth of flax. *Plant Sci* 124: 97–105.
- Cunha AC and Fernandes-Ferreira M (1999) Influence of medium parameters on somatic embryogenesis from hypocotyls explants of flax (*Linum usitatissimum* L.) – Effect of carbon source, total inorganic nitrogen and balance between ionic forms and interaction between calcium and zeatin. *J Plant Physiol* 155: 591–597.
- Cunha AC and Fernandes-Ferreira M (2003) Ontogenic variations in free and esterified fatty acids during somatic embryogenesis of flax (*Linum usitatissimum* L.) *Plant Sci* 164: 863–872.
- Dedicova B, Hricova A, Samaj J, Obert B, Bobak M & Pretova A (2000) Shoots and embryo-like structures regenerated from cultured flax (*Linum usitatissimum* L.) hypocotyl segments. *J Plant Physiol* 157: 327–334.
- Dong JZ and McHughen A (1993) An improved procedure for production of transgenic flax plants using *Agrobacterium tumefaciens*. *Plant Sci* 88: 61–71.
- Erdelska O, Kobeticova D and Pretova A (1973) The *in vitro* development of excised flax embryos. *Biologia* 28: 235–239.
- Gamborg OL & Shyluk JP (1976) Tissue culture, protoplasts and morphogenesis in flax. *Bot. Gaz.* 137: 301–306
- Ibrahim RK (1971) Media for growth of flax tissue culture. *Can J Bot* 49: 295. Jain P and Rashid A (2001) Stimulation of shoot regeneration on *Linum* hypocotyls segments by thidiazuron and its response to light and calcium. *Biol Plantarum* 44: 611–613.
- Jordan MC and McHughen A (1988a) Glyphosate tolerant flax plants from *Agrobacterium* mediated gene transfer. *Plant Cell Rep* 7: 281–284.
- Liao S and Ibrahim RK (1973) Biochemical differentiation in flax tissue culture – phenolic compounds. *Can J Bot* 51: 820–823.
- Ling HQ and Binding H (1992) Improvement of plant regeneration from *Linum* protoplasts by the induction of somatic embryogenesis. *J Plant Physiol* 139: 422–426. Ling HQ and Binding H (1997) Transformation in protoplast cultures of *Linum usitatissimum* and *L. suffruticosum* mediated with PEG and with *Agrobacterium tumefaciens*. *J Plant Physiol* 151: 479–488.
- Link GKK and Eggera V (1946) Mode, site and time of initiation of hypocotyledonary bud primordia in *Linum usitatissimum* L. *Bot Gaz* 107: 441–454.
- McHughen A and Swartz M (1984) A tissue culture derived salt-tolerant line of flax (*Linum usitatissimum*). *J Plant Physiol* 117: 109–117.
- McSheffrey SA, McHughen A and Devine MD (1992) Characterization of transgenic sulfonyleurea-resistant flax (*Linum usitatissimum*). *Theor Appl Genet* 84: 480–486.
- Mlynarova L, Bauer M, Nap JP and Pretova A (1994) High efficiency *Agrobacterium* – mediated gene transfer to flax. *Plant Cell Rep* 13: 282–285.
- Obert B, Benson EE, Millam S, Pretova A and Bremner D (2004b) Moderation of morphogenetic and oxidative stress responses in flax *in vitro* cultures by hydroxynonenal and desferrioxamine. *J Plant Physiol* 162: 537–47.
- O’Connor BJ, Robertson AJ and Gusta LV (1991) Differential stress tolerance and cross adaptation in a somaclonal variant of flax. *J Plant Physiol* 139: 32–36.
- Pretova A (1986) Influence of kinetin on the growth of zygotic flax embryos *in vitro*. *Plant Cell Rep* 3: 210–211.
- Pretova A & Williams EG (1986) Direct somatic embryogenesis from immature zygotic embryos of flax (*Linum usitatissimum* L.). *J Plant Physiol* 126: 155–161.
- Pretova A and Obert B (2003) Flax (*Linum usitatissimum* L.) – A plant system for study of embryogenesis. *Acta Biol Cracov Bot* 45: 15–18.
- Rakousky S, Tejklova E, Wiesner I, Wiesnerova D, Kocabek T and Ondrej M (1999) Hygromycin B – an alternative in flax transformant selection. *Biol Plantarum* 42: 361–369.

- Tejavathi DH, Sita GL and Sunita AT (2000) Somatic embryogenesis in flax. *Plant Cell Tiss Org* 63: 155–159.
- Turel FLM and Ledingham GA (1957) Production of aerial mycelium and uredospores by *Melampsora lini* (Pers) H. Lev on flax leaves in tissue culture. *Can J Microbiol* 3: 813.
- Vromans J (2006) Molecular genetic studies in flax (*Linum usitatissimum* L.). PhD thesis ISBN 90-8504-374-3.
- Wijayanto T and McHughen A (1999) Genetic transformation of *Linum* by particle bombardment. *In Vitro Cell Dev-Pl* 35: 456–465.

### ***Brassica carinata***

- Alemayehu N and Becker H (2002) Genotypic diversity and patterns of variation in a germplasm material of Ethiopian mustard (*Brassica carinata* A. Braun). *Genet Resour Crop Ev* 49: 573-582.
- Babic V, Dalta RS, Scoles GJ and Keller WA (1998) Development of an efficient *Agrobacterium*-mediated transformation system for *Brassica carinata*. *Plant Cell Rep* 17: 183–188.
- Barro F, Escobar J, De la Vega M and Martin A (2002) Modification of glucosinolate and erucic acid contents in doubled haploid lines of *Brassica carinata* by UV treatment of isolated microspores. *Euphytica* 129: 1–6.
- Becker HC, Loeptien H and Roebbelen G (1999) Breeding: an overview. In: C. Gomez-Campo (ed.), *Biology of Brassica Coeno- species*, 413-460. Elsevier, Science BV, Amsterdam.
- Chaudhary S, Parmenter DL and Moloney MM (1997) Transgenic *Brassica carinata* as a vehicle for the production of recombinant proteins in seeds. *Plant Cell Rep* 17: 195–200.
- Choung PV, Pauls KP and Beversdorf WD (1987) Protoplast culture and plant regeneration from *Brassica carinata* Braun. *Plant Cell Rep* 6: 67-69.
- Downey RK and Röbbelen G (1989) *Brassica* species. In: Röbbelen G, Downey RK, Ashri A (eds) *Oil crops of the world*. McGraw-Hill, New York, pp 342–344.
- Downey R K (1990) Canola: a quality Brassica oilseed. In: Janick, J. and J. E. Simon (eds), *Advances in New Crops*, 211-217. Timber Press, Portland.
- Gupta V, Agnihotri A and Jagannathan V (1991) Plant regeneration from callus and protoplasts of *Brassica nigra* through somatic embryogenesis. *Plant Cell Rep* 9: 427-430.
- Hasterok R and Maluszynska J (2000a) Cytogenetic markers of *Brassica napus* chromosomes. *J Appl Genet* 41: 1–9.
- Hasterok R and Maluszynska J (2000b) Cytogenetic analysis of diploid *Brassica* species. *Acta Biol Cracov Ser Bot* 42: 145–163.
- Hasterok R and Maluszynska J (2000c) Nucleolar dominance does not occur in root tip cells of allotetraploid *Brassica* species. *Genome* 43: 574–579.
- Jaiswal SK, Hammatt N, Bhojwani SS, Cocking EC and Davey MR (1990) Plant regeneration from cotyledon protoplasts of *Brassica carinata*. *Plant Cell Tiss Org Culture* 22: 159-165.
- Kao HM, Keller WA, Gleddie S and Brown GG (1990) Efficient plant regeneration from hypocotyl protoplasts of broccoli (*B. oleracea* L. *ssp. Italica Planck*). *Plant Cell Rep* 9: 311-315.
- Kinney AJ, Cahoon EB and Hitz WD (2002) Manipulating desaturase activities in transgenic crop plants. *Biochem. Soc. Trans.* 30: 1099–1103.
- Kirti PB and Chopra VL (1990) Rapid plant regeneration through organogenesis and somatic embryogenesis from cultured protoplasts. *Plant Cell Tiss Org* 20: 65- 67.
- Malik RS (1990) Prospects for *Brassica carinata* as an oilseed crop in India. *Exp. Agric.* 26: 125-129.
- Maluszynski M, Ahloowalia BS and Sigurbjörnsson B (1995) Application of *in vitro* mutation techniques for crop improvement. *Euphytica* 85: 303–315.
- Narasimhulu SB, Kirti PB, Prakash S and Chopra VL (1992) Rapid and efficient plant regeneration from hypocotyl protoplasts of *Brassica carinata*. *Plant Cell Rep* 11:159-162.
- Poulsen GB (1995) Genetic transformation of Brassica. *Plant breeding* 115: 209- 225.
- Raney P, Rakow G and Olson T (1995) Modification of Brassica seed oil fatty acid composition utilizing interspecific crossing. In: D. Murphy (ed.), *Proc. 9th Int. Rapeseed Conf.*, Cambridge, 4-7 July 1995, 410-412. GCIRC, Cambridge.

- Sheikh FA, Lone B, Najeeb S, Shikari AB, Parray GA, Rather AG and Khudwani RS (2009) Induced mutagenesis for seed quality traits in Ethiopian mustard (*Brassica carinata a. braun*). ARPN Journal of Agricultural and Biological Science, vol 4.
- Thomzik JE and Hain R (1990) Transgenic *Brassica napus* plants obtained by cocultivation of protoplasts with *Agrobacterium tumefaciens*. Plant Cell Rep 9: 233-236.
- Velasco L, Fernandez-Martinez J and De Haro A (1995a) Isolation of induced mutants in Ethiopian mustard (*Brassica carinata Braun*) with low levels of erucic acid. Plant Breeding 114: 454-456.
- Velasco L, Fernandez-Martinez J and De Haro A (1998) Increasing erucic acid content in Ethiopian mustard through mutation breeding. Plant Breeding 117, 85—87.
- Yang M-Z and Jia S-R (1989) Plant regeneration from protoplasts of *Brassica carinata* Braun. Acta Bot Sin 31: 89-94.

### ***Brassica napus***

- Barfield DG and Pua EC (1991) Gene transfer in plants of *Brassica juncea* using *Agrobacterium tumefaciens* mediated transformation. Plant Cell Rep. 10: 308-314.
- Burstin J, Marget P, Huart M, Moessner A, Mangin B et al (2007) Developmental genes have pleiotropic effects on plant morphology and source capacity, eventually impacting on seed protein content and productivity in pea. Plant Physiol. 144: 768-781.
- De Block M, De Brower D and Tenning P (1989) Transformation of *Brassica napus* and *Brassica oleracea* using *Agrobacterium tumefaciens* and the expression of the bar and neo genes in the transgenic plants. Plant Physiol 91: 694-701.
- Eapen S and George L (1997) Plant regeneration from peduncle segments of oil seed Brassica species: influence of silver nitrate and silver thiosulfate. Plant Cell Tiss. Org 51: 229-232.
- Gupta V, Sita GL, Shaila MS and Jagannathan V (1993) Genetic transformation of *Brassica nigra* by *Agrobacterium* based vector and direct plasmid uptake. Plant Cell Rep. 12: 418-421.
- Hachey JE, Sharma KK and Moloney MM. (1991) Efficient shoot regeneration of *Brassica campestris* using cotyledon explants cultured *in vitro*. Plant Cell Rep 9: 549-554.
- Halfhill MD, Richards HA, Mabon SA and Stewart CN Jr (2001) Expression of GFP and Bt transgenes in *Brassica napus* and hybridization with *Brassica rapa*. Theor Appl Genet 103: 659-667.
- Hochholdinger F, Wen TJ, Zimmermann R, Chimot- Marolle P, da Costa e Silva O et al. (2008) The maize (*Zea mays* L.) roothairless 3 gene encodes a putative GPI-anchored, monocot-specific, COBRA-like protein that significantly affects grain yield. Plant J 54: 888-898.
- Hu Q, Anderson SB and Hansen L (1999) Plant regeneration capacity of mesophyll protoplasts from *Brassica napus* and related species. Plant Cell Tiss Org 59: 189-196.
- Jagannath A, Arumugam N, Gupta V, Pradhan A, Burma PK and Pental D. (2002) Development of transgenic barstar lines and identification of a male sterile (barnase)/restorer (barstar) combination for heterosis breeding in Indian oilseed mustard (*Brassica juncea*). Curr Sci 82: 46-52.
- Kirti PB, Prakash S and Chopra VL (1991b) Interspecific hybridization between *Brassica juncea* and *B. spinescens* through protoplast fusion. Plant Cell Rep 9: 639-642.
- Klimaszewska K and Keller K (1985) High frequency plant regeneration from thin cell layer explants of *Brassica napus*. Plant Cell Tiss Org 4: 183-197.
- Leroy XJ, Leon K, Charles G and Branchard M (2000) Cauliflower somatic embryogenesis and analysis of regenerant stability by ISSRs. Plant Cell Rep 19: 1102-1107.
- Leroy XJ, Leon K, Hily JM, Chaumeil P and Branchard M (2001) Detection of *in vitro* culture-induced instability through inter-simple sequence repeat analysis. Theor Appl Genet 102: 885-891.
- Liu JW, DeMichele S, Bergana M, Bobik E, Hastilow C, Chuang LT, Mukerji P and Huang YS (2001) Characterization of oil exhibiting high gamma-linolenic acid from a genetically transformed canola strain. J Am Oil Chem Soc 78: 489-493.
- Moloney MM, Walker JM and Sharma KK (1989) High efficiency transformation of *Brassica napus* using *Agrobacterium* vectors. Plant Cell Rep 8: 238-242.
- Ono Y, Takahata Y and Kaizuma N (1994) Effect of genotype on shoot regeneration from cotyledonary explants of rapeseed (*Brassica napus* L.). Plant Cell Rep 14: 13-17.
- Pan J, Wang J, Zhou Z, Yan Y, Zhang W, Lu W, Ping S, Dai Q, Yuan M, Feng B, Hou X, Zhang Y, Ma R, Liu T, Feng L, Wang L, Chen M and Lin M (2009) IrrE a global regulator of



- extreme radiation resistance in *Deinococcus radiodurans*, enhances salt tolerance in *Escherichia coli* and *Brassica napus*. Plos One 4, e4422.
- Radke SE, Turner JC and Facciotti D (1992) Transformation and regeneration of *Brassica rapa* using *Agrobacterium tumefaciens*. Plant Cell Rep 11: 499–505.
  - Ren JP, Dickson MH and Earle ED (2000) Improved resistance to bacterial soft rot by protoplast fusion between *Brassica rapa* and *B. oleracea*. Theor Appl Genet 100: 810–819.
  - Shomura A, Izawa T, Ebana K, Ebitani T, Kanegae H et al. (2008) Deletion in a gene associated with grain size increased yields during rice domestication. Nat Genet 40 1023–1028.
  - Turgut K, Barghchi M and Scott (1998) Efficient shoot regeneration and somatic embryogenesis from immature cotyledons of *Brassica napus* L. Plant Breed 117: 503–504.
  - Wang WC, Menon G and Hansen G (2003) Development of a novel *Agrobacterium* mediated transformation method to recover transgenic *Brassica napus* plants. Plant Cell Rep 22: 274–281
  - Warrington CV, Zhu S, Parrott WA, All JN and Boerma HR (2008) Seed yield of near-isogenic soybean lines with introgressed quantitative trait loci conditioning resistance to corn earworm (Lepidoptera: Noctuidae) and soybean looper (Lepidoptera: Noctuidae) from PI 229358. J Econ Entomol 101: 1471–1477.
  - Xie X, Jin F, Song MH, Suh JP, Hwang HG et al. (2008) Fine mapping of a yield-enhancing QTL cluster associated with transgressive variation in an *Oryza sativa* and *O. rufipogon* cross. Theor Appl Genet 116: 613–622.
  - Xing YZ, Tang WJ, Xue WY, Xu CG and Zhang Q (2008) Fine mapping of a major quantitative trait loci, qSSP7, controlling the number of spikelets per panicle as a single Mendelian factor in rice. Theor Appl Genet 116: 789–796.
  - Xu ZH, Davey MR and Cocking EC (1982) Plant regeneration from root protoplasts of *Brassica*. Plant Sci Lett 24: 117–121.
  - Xue W, Xing Y, Weng X, Zhao Y, Tang W et al. (2008) Natural variation in *Ghd7* is an important regulator of heading date and yield potential in rice. Nat Genet 40: 761–767.
  - Yang MZ, Jia SR and Pua EC (1991) High frequency of plant regeneration from hypocotyl explants of *Brassica carinata* A.Br. Plant Cell Tiss Org 24: 79–82.

### ***Miscanthus spp***

- Andersson NJ (1855) Om de med Saccharum beslägtade genera. Öfvers. Kunglia VetenskapsAkademienFörn.,Stockholm 12:151-167.
- Baldwin BG (1992) Phylogenetic utility of the internal transcribed spacers of nuclear ribosomal DNA in plants: An example from the Compositae. Mol. Phylogen. Evolution, 1: 3–16.
- Beale CV and Long SP (1995) Can perennial C4 grasses attain high efficiencies of radiant energy conversion in cool climates? Plant, Cell & Environment 18: 641–650.
- Cappelletto P, Mongardini F, Barberi B, Sannibale M, Brizzi M and Pignatelli V (2000) Papermaking pulps from the fibrous fraction of *Miscanthus x giganteus*. Industrial Crops and Products 11: 205–210.
- Chaw SM, Zharkikh A, Sung HM, Lau TC and Li WH (1997) Molecular phylogeny of extant gymnosperms and seed plant evolution: Analysis of nuclear 18S rRNA sequences. Mol Biol Evol.14: 56–68.
- Deuter M and Abraham J (1998) Genetic resources of *Miscanthus* and their use in breeding. In: Kopetz H, Weber T, Palz W, Chartier P, Ferrero GL, eds. Biomass for energy and industry. Rimpf, Germany: C.A.R.M.E.N.: 775–777.
- Greef MJ, Deuter M, Jung C, Schondelmaier J (1997) Genetic diversity of European *Miscanthus* species revealed by AFLP fingerprinting. Genet Res Crop Evol 44:185–195.
- Hernández P, Dorado G, Laurie DA, Martín A and Snape JW (2001) Microsatellites and RFLP probes from maize are efficient sources of molecular markers for the biomass energy crop *Miscanthus*. Theor Appl Genet 102:616–622.
- Hodkinson TR and Renvoize S (2001) Nomenclature of *Miscanthus x giganteus* (*Poaceae*). Kew Bulletin 56: 759–760.
- Holme IB and Petersen KK (1996) Callus induction and plant regeneration from different explant types of *Miscanthus x ogiformis* Honda Giganteus. Plant Cell Tiss Org 45: 43–52.
- Kjeldsen, JB, Jørgensen U and Kristensen EF (1999) *Miscanthus* for thatching. In: Markbrug. DJF Rapport, Foulum: p. 59.
- Lewandowski I, Clifton-Brown JC, Scurlock JMO and Huisman W (2000) *Miscanthus*: European experience with a novel energy crop. Biomass Bioenerg 19: 209–277.

- Lewandowski I and Kahnt G (1993) Möglichkeiten zur Erstellung eines *In-vitro*-Vermehrungssystems für *Miscanthus Giganteus* durch Nutzung verschiedener Pflanzenteile als Spendergewebe. *Bodenkultur* 44: 243–252.
- Nielsen PN (1987) Vegetativ formering af elefantgræs, *Miscanthus sinensis* Giganteus. *Tidsskr. Planteavl* 91: 361–368.
- Nielsen JM, Brandt K and Hansen J (1993) Long-term effects of thidiazuron are intermediate between benzyladenine, kinetin or isopentenyladenine in *Miscanthus sinensis*. *Plant Cell Tiss Org* 35: 173–179.
- Parsby M (1996) Straw and Energy Crops – Analyses of Economy, Energy and Environment (in Danish). Statens Jordbrugs- og Fiskeriøkonomiske Institut. Report no. 87.
- Petersen KK (1997) Callus induction and plant regeneration in *Miscanthus x ogiformis* Honda ‘Giganteus’ as influenced by benzyladenine. *Plant Cell Tiss Org* 49: 137–140.
- Venendaal R, Jørgensen U and Foster CA (1997) European energy crops: a synthesis. *Biomass Bioenergy* 13:147–185.
- Wagenaar BM and Vandenheuvel EJMT (1997) Co-combustion of *Miscanthus* in a pulverised coal combustor—experiments in a drop tube furnace. *Biomass Bioenergy* 12:185–197.
- Wang D, Portis Jr. AR, Moose SP and Long SP (2008) Cool C4 Photosynthesis: Pyruvate Pi Dikinase Expression and Activity Corresponds to the Exceptional Cold Tolerance of Carbon Assimilation in *Miscanthus x giganteus*. *Plant Physiol* 148:557–567.

### *Panicum virgatum*

- Alexandrova KS, Denchev PD and Conger BV (1996). In vitro development of inflorescences from switchgrass nodal segments. *Crop Sci* 36: 175–178.
- Carroll A and Somerville C (2009) Cellulosic Biofuels. *Annu Rev Plant Biol* 60:165-182.
- Casler MD, Stendal CA, Kapich L and Vogel KP (2007) Genetic Diversity, Plant Adaptation Regions, and Gene Pools for Switchgrass. *Crop Sci* 47: 2261 - 2273.
- Hultquist SJ, Vogel KP, Lee DJ, Arumuganathan K and Kaeppler S (1996) Chloroplast DNA and nuclear DNA content variations among cultivars of switchgrass, *Panicum virgatum* L. *Crop Sci* 36:1049–1052.
- Li J-F, Park E, von Arnim AG and Nebenführ A (2009) The FAST technique: a simplified *Agrobacterium*-based transformation method for transient gene expression analysis in seedlings of Arabidopsis and other plant species. *Plant Methods* 5: 6.
- Lu, K, Kaeppler SM, Vogel KP, Arumuganathan K and Lee DJ (1998) Nuclear DNA content and chromosome numbers in switchgrass. *Great Plains Res* 8: 269–280.
- Martinez-Reyna JM. and Vogel KP (2008) Heterosis in Switchgrass: Spaced Plants. *Crop Sci* 48: 1312–1320
- Mazarei M, Al-Ahmad H, Rudis MR and Stewart CN (2008) Protoplast isolation and transient gene expression in switchgrass, *Panicum virgatum* L. *Biotechnol J* 3: 354-359.
- McLaughlin SB and Kszos LA (2005) Development of switchgrass (*Panicum virgatum*) as a bioenergy feedstock in the United States. *Biomass Bioenergy* 28: 515-535.
- Missaoui A.M., Paterson A.H. and Bouton J.H. (2005) Investigation of genomic organization in switchgrass (*Panicum virgatum* L.) using DNA markers. *Theor Appl Genet* 110: 1372–1383.
- Richards HA, Rudas VA, Sun H, McDaniel JK, Tomaszewski Z and Conger BV (2001) Construction of a GFP-BAR plasmid and its use for switchgrass transformation. *Plant Cell Reports* 20: 48–54.
- Saha MC, Chekhovskiy K, Narasimhamoorthy B and Bouton JH (2007) Cross-species amplification of microsatellite markers in switchgrass. p. 75. Proc. of 29th Symposium on Biotechnology for Fuels and Chemicals, 29 April–2 May 2007, Denver, CO ([http://www.simhq.org/meetings/29symp/29fuelsdraft\\_WEB.pdf](http://www.simhq.org/meetings/29symp/29fuelsdraft_WEB.pdf))
- Somleva MN, Snell KD, Beaulieu JJ, Peoples OP, Garrison BR and Patterson NA (2008) Production of polyhydroxybutyrate in switchgrass, a value-added co-product in an important lignocellulosic biomass crop. *Plant Biotechnol J* 6: 663-678 .
- Tobias CM, Sarath G, Twigg P, Lindquist E, Pangilinan J, Penning BW, Barry K, McCann MC, Carpita NC and Lazo GR (2008) Comparative Genomics in Switchgrass Using 61,585 High-Quality Expressed Sequence Tags. *Plant Genome* 1: 111–124.
- Vogel KP and Mitchell R (2007) Heterosis in switchgrass: Biomass yield in swards. *Crop Sci* 48: 2159-2164.